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# Valorization of Natural Tomato Wastes in the Reduction of Prochiral Ketones

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## ABSTRACT

In recent years, a great amount of attention has been paid to asymmetric synthesis of chiral synthons, the demand for which is increasing as precursors in the development of modern drugs and agrochemicals. Chiral alcohols are one of the many well-known synthons and can be obtained from the corresponding prochiral ketones by asymmetric reduction.. In this work,Organic chemical bioconversion of prochiral ketones to chiral alcohols with natural tomatoes food waste was made in water and glycerol and compared to known strains of waste and microorganisms (*e.g. Baker's Yeast , Daucus carota*). These enzymatic reduction attempts lead to best results with excellent enantiomeric excess (ee) in *S* configuration.

Keywords: Tomato wastes, enzymatic reduction, secondary alcohols, enantioselectivity.



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#### INTRODUCTION

The production and the preservation of the environment without the alteration of the soil and water space during the production or manufacture should be the main condition and one of the greatest preoccupation of public administration.[1] In the East of Algeria, many tomato processing factories are installed around Seybouse Valley and produce a lot of food wastes, source of soil and water pollution of this fertile agricultural plain. In addition to their use as a fertilizer of natural composts, biogas ... etc.[2–4] these wastes can be recycled as valuable organic products with many features of bioconversion in organic and green chemistry and other uses. [5, 6]. It is well know that vegetable and fruit processing by-products, wastes and effluents typically consist of high amounts of proteins, sugars and lipids[7, 8] can be sources of enzyme activity and can be used as a biocatalyst product in various organic chemistry reactions.[9, 10]

On the other hand, chemical synthesis of organic molecules with chiral centers often gives a racemic mixture that is difficult and expensive to separate, presenting a major obstacle to accede to many fine active biologically chemical products. Among the many types of biocatalysts used, the carbonyl reductases were used for preparing optically active alcohols from various prochiral ketones [11–14] For example, in organic synthesis, the reduction of a prochiral ketone to the secondary alcohol, [15–17] is an enantioselective manner among others that could be explored in this valorization. [18, 19].



## Figure 1 Prochiral ketone reduction

In particular, when the enantioselectivity of the alcohol produced is very high, [20–22] and has a great interest as building block [23, 24] for accede to potentially biologically active products. [25, 26]

## MATERIELS AND METHODS

Tomato wastes are freshly collected in plant manufacture [59] and are directly used without any processing in same day. The reduction's attempts with tomato fruit [60] bought to local market and used after having been cut into small slices are best and comparable with tomato wastes. The conversion and products ratio is determined by GLC.

The enantiomeric composition of product was established by chiral column FS Cyclodex beta- I/P over Shimatzu GC 4010 and P1000-LED polarimeter with precision of lecture 0.05°. Configuration of product was determined by the sign of the specific rotation and in accordance with literature. [61, 62]

## EXPERIMENTAL

## A general procedure used for Bioreduction with

Tomato wastes are in accordance with reference cited [53]. The results are detailed in the table. Tomatoes wastes (5 g), or a tomato cut into small dices (50g) was placed in Erlenmeyer flasks with 50 mL of solvent (water or glycerol)[54]. (50 mg) of the substrate [55] was added to the suspension of the tomato, and the resulting mixture was agitated by a mechanical stirrer with the agitation speed at 300 rpm. The temperature of the reaction system was kept at 30 °C. The process of the reaction was monitored by GC and comparison with product reduces with chemical method. [56, 57, 58]

March – April

2017

RJPBCS

8(2) Page No. 121



After completion of the reaction, the suspension was filtered off and the filtrate was extracted with ethyl acetate ((3x40 mL for water and 3x20 mL for glycerol); then the combined organic layer was dried over anhydrous MgSO4 and concentrated in vacuo. The crude mixture was purified by silica gel column chromatography (dichloromethane: methanol = 9:1).

### **RESULTS AND DISCUSSION**

The aim of this work was to make several attempts with raw food waste on various prochiral ketones, to know the reduction capacity of waste, optimize the reaction conditions and the solvent use.[27–29] Our interest for tomato waste is dictated by the lack of serious and comprehensive studies of this plant in the field of enzyme activity such as reductase.[30] For these BioReagents / catalysts widely applied in synthetic chemistry, they need to operate in retaining their selectivity in more compatible with organic compounds and solvents. For example: the Baker's yeast is known to perform a variety of transformations in synthetic organic chemistry, such as reduction of carbonyl groups.

However, its use in this field has been limited by the necessity of employing aqueous solvent systems. They exist several works related to these problems. [31, 32] The main advantage of using organic solvents is the ease with which the product can be extracted. Alternative biocatalyst research with the same activity among domestic waste, could be an interesting approach. In this paper, we are particularly interested in the tomato activity (Lycopersicum esculentum). So, we studied the tomato reduction activity in both solvents (water and glycerol). The choice of substrate is made by disposability and the structure of prochiral linear ketone : octan-3-one (III) and butan-2-one (V), aromatic cycle: acetophenone (I), aromatic cycle with heteroatome: p-nitroacetophenone (IV) 3-chloro-1-phenylpropan-1- one (VI), and aromatic cycle with substituted alkyl: 4-methylacetophenone (II). The Table 1 below shows some attempts of reduction with Tomato (T) over seven prochiral ketones cited before. The global results of these attempts, shows that enantiomeric excesses in water or glycerol as solvent are the best and (S) conversion is according with Prelog's rule. results.

Na	Red. Sub.		Conv.% t(h)	ee (%)		Conf.	
			Wat. Gly.	Wat. Gly.		Wat. Gly.	
1	Т	Ι	40(96) 89(96)	> 99	> 99	S	S
2	T.	Π	40(96) 87(96)	>99	>99	S	S
3	T.	ш	< 2(180) 12(96)	-	> 99	-	S
4	T.	IV	36(96) 29(96)	>99	> 99	S	S
5	T.	V	< 2(180) < 3(96)	-	-	-	_
6	T.	VI	100(72) < 1(120)	> 99	>99	S	s

#### Table 1 The reduction tests with Tomatoes wastes on prochiral ketones in water and glycerol.

Acetophenone (I), 4-methylacetophenone ((II)),Octan-3-one (III), p-nitroacetophenone (IV), butan-2-one (V), 3-chloro-1-phenylpropan-1-one (VI).

#### Reactivity

All the essays realised for understanding various aspects of the reactivity in the enzymatic pocket [33, 34], according to the nature of the substituents R1 and R2 of the substrate (see Fig.1), and explain if either the volume or electronic character or the both can influence reactivity and selectivity. So, if the selectivity does not alter in any case, the reaction kinetic were affected by structure and nature of the substrate; we think, if the reaction is slow with aliphatic compounds, 3-octanone (II) and butan-2-one (V) this is mainly due to the fact, that this kind of products is not well accepted by the enzyme site. In this case, we believe that this kind of



substrate - a prochiral ketone - with apolar substituent R1 and R2 isn't adapted with residues of the enzymatic receptor pocket, which tends to accept the aromatic and polar patterns. For example, in the case where R2 is chloroethyl: a polar group, the reduction yield is almost quantitative ( see entry 6 in Table 1) after just two to three days. However, in the case where the aromatic substituent have the donors or withdrawing groups in para position, this does not alter significantly the kinetic reaction as for example when the moieties in para on the benzene are methyl or a nitro groups. [35,36, 37] In general, we can say that the kinetic reaction is rather dictated by electronics aspects than the steric considerations related to the volume of substituent. However we could not understand the slow reactivity of chlorinated substrate VI in glycerol ( see entry 6 in Table 1) compared with the same essay carried out in the water.[38]. Finally, we will say, that it is difficult to understand all aspects of one complex system where many variables are involved, but understand the essential. This is understandable and consistent with the literature.[39, 40].

When the reaction is finished [41], the extraction of product from water with an organic solvent requires large amounts of solvent [42]. This method is a very expensive process. However with the glycerol the extraction is simpler and the amounts of solvent are less than with the water extraction [43]. This is reflected on the conversion that is better with the glycerol, but with the same enantioselectivity. The dilemma in this manipulation is a choice of solvent: the water is inexpensive and abundant compared to glycerol where the extraction conditions are better. [44–46]



Figure 2 hydrure attack on re-face with prelog's rule.

## Stereoselectivity

To understand the stereochemical trend in the enzymatic reductions [47, 48], all the products in Table 1 are drawn in such a way that the \_-face (*re-face*) of the carbonyl group in the substrates is attacked by H– (NADPH) (see fig 2). The enantiomeric purity of these products was high in most cases, where the substrate is in desired structure and the Prelog's rule is confirmed (entry 1 and 4 in Table 1). Although we have previously reported that the present reductase is one of the enzymes that contribute to Prelogs rule[49, 50], the anti-Prelog-type[51, 52] alcohols are not observed in this study; the left-hand moiety is bulkier than the right-hand moiety. In second attempt the glycerol (Table 1) is used and the results are reproduced below. The assignment of the *R* and *S* configuration has been confirmed by the optical rotation and chiral chromatography. We have noted that the *R* product is more retained than the *S* product in the all attempts. This confirms the attack of the hydrure on the side Re.

## CONCLUSION

In this work we wanted to show the activity of tomato (*Lycopersicum esculentum*). That is interesting for valorization as biotechnological process for synthesis biological active secondary alcohols with *S* configuration.

March - April

2017

RJPBCS

8(2)

Page No. 123



If this activity is slow compared with another reductases, [63] in particular with the small prochiral ketones, when the enzymatic pocket can't accept this substrates. We could show that the selectivity is always excellent when the reaction works and respect Prelog's rules, even if it is slow. We were able to define the structure and nature of the substituents around the carbonyl, for the best yields. Thus the best substrate is one with polar substituents. We believe that the matter responsible of enzymatic activity is not very significant compared to the total mass and can be adjusted by the large quantity of tomato wastes. We have also shown that the use of glycerol can increase the yield of the reaction without being very significant. Finally we have also shown that the S-enantiomer is more retained by the chiral chromatography column and leaves the latter in all cases studied.

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## REFERENCES

- [1] P. J. Withers, J. J. Elser, J. Hilton, H. Ohtake, W. J. Schipper, K. C. van Dijk, Greening the global phosphorus cycle: how green chemistry can help achieve planetary p sustainability, Green Chemistry 17 (4) (2015) 2087–2099.
- [2] J. H. Clark, F. Deswarte, Introduction to chemicals from biomass, John Wiley & Sons, 2015. [3] Y. L. Chiew, J. Spångberg, A. Baky, P.-A. Hansson, H. J<sup>-</sup>onsson, Environmental impact of recycling digested food waste as a fertilizer in agriculturea case study, Resources, Conservation and Recycling 95 (2015) 1–14.
- [3] Y. L. Chiew, J. Spångberg, A. Baky, P.-A. Hansson, H. J¨onsson, Environmental impact of recycling digested food waste as a fertilizer in agriculturea case study, Resources, Conservation and Recycling 95 (2015) 1–14.
- [4] F. Fava, G. Totaro, L. Diels, M. Reis, J. Duarte, O. B. Carioca, H. M. Poggi-Varaldo, B. S. Ferreira, Biowaste biorefinery in europe: opportunities and research & development needs, New biotechnology 32 (1) (2015) 100–108.
- [5] T. Matsuda, R. Yamanaka, K. Nakamura, Recent progress in biocatalysis for asymmetric oxidation and reduction, Tetrahedron: Asymmetry 20 (5) (2009) 513 557.
- [6] K. Faber, Biotransformations in organic chemistry: a textbook, Springer Science & Business Media, 2011.
- [7] A. Moayedi, M. Hashemi, M. Safari, Valorization of tomato waste proteins through production of antioxidant and antibacterial hydrolysates by proteolytic bacillus subtilis: optimization of fermentation conditions, Journal of Food Science and Technology (2015) 1–10And reference cited.
- [8] G. Boschin, G. M. Scigliuolo, D. Resta, A. Arnoldi, Aceinhibitory activity of enzymatic protein hydrolysates from lupin and other legumes, Food chemistry 145 (2014) 34–40.
- [9] K. Drauz, Enzyme catalysis in organic synthesis: a comprehensive handbook, Vol. 1, John Wiley & Sons, 2012.
- [10] M. T. Reetz, Biocatalysis in organic chemistry and biotechnology: past, present, and future, Journal of the American Chemical Society 135 (34) (2013) 12480–12496.
- [11] Y. Ni, J.-H. Xu, Biocatalytic ketone reduction: A green and efficient access to enantiopure alcohols, Biotechnology Advances 30 (6) (2012) 1279 – 1288.
- [12] T. S. Moody, J. D. Rozzell, Chapter 6 modern biocatalytic ketone reduction, in: A. G. D. Stewart (Ed.), Organic Synthesis Using Biocatalysis, Academic Press, 2016, pp. 149–185.
- [13] R. L. Cunha, E. A. Ferreira, C. S. Oliveira, 'A. T. Omori, Biocatalysis for desymmetrization and resolution of stereocenters beyond the reactive center: How far is far enough?, Biotechnology advances 33 (5) (2015) 614–623.
- [14] R.Wohlgemuth, Development of sustainable biocatalytic reduction processes for organic chemists, Synthetic Methods for Biologically Active Molecules: Exploring the Potential of Bioreductions (2014) 1– 25.
- [15] F. Hollmann, I. W. Arends, D. Holtmann, Enzymatic reductions for the chemist, Green Chemistry 13 (9) (2011)2285–2314.



- [16] K. Goldberg, K. Schroer, S. L¨utz, A. Liese, Biocatalytic ketone reductiona powerful tool for the production of chiral alcoholsparti: processes with isolated enzymes, Applied microbiology and biotechnology 76 (2) (2007) 237–248.
- [17] I. A. Kaluzna, T. Matsuda, A. K. Sewell, J. D. Stewart, Systematic investigation of saccharomyces cerevisiae enzymes catalyzing carbonyl reductions, Journal of the American Chemical Society 126 (40) (2004) 12827–12832.
- [18] C. M. Galanakis, Recovery of high added-value components from food wastes: Conventional, emerging technologies and commercialized applications, Trends in Food Science Technology 26 (2) (2012) 68 – 87.
- [19] A. A. Koutinas, A. Vlysidis, D. Pleissner, N. Kopsahelis, I. L. Garcia, I. K. Kookos, S. Papanikolaou, T. H. Kwan, C. S. K. Lin, Valorization of industrial waste and by-product streams via fermentation for the production of chemicals and biopolymers, Chemical Society Reviews 43 (8) (2014) 2587–2627.
- [20] C. J. Sih, S.-H. Wu, Resolution of enantiomers via biocatalysis, Topics Stereochem 19 (1989) 63–125.
- [21] J. Rakels, A. Straathof, J. Heijnen, A simple method to determine the enantiomeric ratio in enantioselective biocatalysis, Enzyme and Microbial Technology 15 (12) (1993) 1051 1056.
- [22] K. Shin-ya, H. Sugeta, S. Shin, Y. Hamada, Y. Katsumoto,K. Ohno, Absolute configuration and conformation analysis of 1-phenylethanol by matrix-isolation infrared and vibrational circular dichroism spectroscopy combined with density functional theory calculation, The Journal of Physical Chemistry A 111 (35) (2007) 8598–8605.
- [23] H. H. Szmant, Organic building blocks of the chemical industry, John Wiley & Sons, 1989.
- [24] E. Santaniello, P. Ferraboschi, P. Grisenti, A. Manzocchi, The biocatalytic approach to the preparation of enantiomerically pure chiral building blocks, Chemical reviews 92 (5) (1992) 1071–1140.
- [25] A. Harvey, Strategies for discovering drugs from previouslyunexplored natural products, Drug discovery today 5 (7) (2000) 294–300.
- [26] K. M. Koeller, C.-H. Wong, Enzymes for chemical synthesis, Nature 409 (6817) (2001) 232–240.
- [27] A. Zaks, A. M. Klibanov, The effect of water on enzyme action in organic media., Journal of Biological Chemistry 263 (17) (1988) 8017–8021. Resources, Conservation and Recycling 95 (2015) 1 14.
- [28] R. V. Rariy, A. M. Klibanov, Correct protein folding in glycerol, Proceedings of the National Academy of Sciences 94 (25) (1997) 13520–13523.
- [29] W. Kroutil, H. Mang, K. Edegger, K. Faber, Recent advances in the biocatalytic reduction of ketones and oxidation of secalcohols, Current opinion in chemical biology 8 (2) (2004) 120–126.
- [30] K. Phukan, N. Devi, Biocatalytic preparative method of asymmetric alcohols using lycopersicum esculentum (tomato), Int J Chem Tech Research 4 (2012) 203–207.
- [31] M. K. Johns, A. J. Smallridge, M. A. Trewhella, The use of liquefied petroleum gas (lpg) as a solvent for yeast reactions, Tetrahedron Letters 42 (25) (2001) 4261–4262.
- [32] L. Y. Jayasinghe, A. J. Smallridge, M. A. Trewhella, The yeast mediated reduction of ethyl acetoacetate in petroleum ether, Tetrahedron letters 34 (24) (1993) 3949–3950.
- [33] I. Castellano, A. Merlino, -glutamyltranspeptidases: sequence, structure, biochemical properties, and biotechnological applications, Cellular and Molecular Life Sciences 69 (20) (2012) 3381–3394.
- [34] J. Uppenberg, N. Oehrner, M. Norin, K. Hult, G. J. Kleywegt, S. Patkar, V. Waagen, T. Anthonsen, T. A. Jones, Crystallographic and molecular-modeling studies of lipase b from candida antarctica reveal a stereospecificity pocket for secondary alcohols, Biochemistry 34 (51) (1995) 16838–16851.
- [35] In the case where the group is hydroxyl, the reaction is slow and not significant and confirm than the acidity of phenol moiety can perturb enzymatic reaction. This attempt was abandoned because not interesting.
- [36] A. I. Ca<sup>~</sup>nas, S. Camarero, Laccases and their natural mediators: biotechnological tools for sustainable eco-friendly processes, Biotechnology advances 28 (6) (2010) 694–705.
- [37] R.McCague, M. G. Rowlands, S. E. Barrie, J. Houghton, Inhibition of enzymes of estrogen and androgen biosynthesis by esters of 4-pyridylacetic acid, Journal of medicinal chemistry 33 (11) (1990) 3050– 3055.
- [38] Several tests were carried out in glycerol with the **VI** substrate: without any success.
- [39] M. Sutter, E. D. Silva, N. Duguet, Y. Raoul, E. Metay, M. Lemaire, Glycerol ether synthesis: A bench test for green chemistry concepts and technologies, Chemical reviews 115 (16) (2015) 8609–8651.
- [40] J. I. Garc'ıa, H. Garc'ıa-Mar'ın, E. Pires, Glycerol based solvents: synthesis, properties and applications, Green Chemistry 16(3) (2014) 1007–1033.
- [41] The reaction is stirred for a minimum of four days.



- [42] One (01) liter of ethylacetate is used for extract 400 mg of product from the mixture and the ethyl acetate used is continually recycled.
- [43] We used the half quantity of ethylacetate when we used the glycerol.
- [44] M. Hall, C. Stueckler, H. Ehammer, E. Pointner, G. Oberdorfer, K. Gruber, B. Hauer, R. Stuermer, W. Kroutil, P. Macheroux, et al., Asymmetric bioreduction of c? c bonds using enoate reductases opr1, opr3 and yqjm: Enzyme-based stereocontrol, Advanced Synthesis & Catalysis 350 (3) (2008) 411–418.
- [45] M. Hall, C. Stueckler, W. Kroutil, P. Macheroux, K. Faber, Asymmetric bioreduction of activated alkenes using cloned 12- oxophytodienoate reductase isoenzymes opr-1 and opr-3 from lycopersicon esculentum (tomato): A striking change of stereoselectivity, Angewandte Chemie 119 (21) (2007) 4008–4011.
- [46] L. Agwaramgbo, T. Okegbe, T. Wright, S. Igwe, V. Ogburie, Inhibition of the oxidation of acetophenone by aqueous extracts of edible fruits and vegetables, Mod Chem appl 1 (105) (2013)2.
- [47] H. S. Toogood, N. S. Scrutton, New developments in enereductase catalysed biological hydrogenations, Current opinion in chemical biology 19 (2014) 107–115.
- [48] M. L. Contente, I. Serra, M. Brambilla, I. Eberini, E. Gianazza, V. De Vitis, F. Molinari, P. Zambelli, D. Romano, Stereoselective reduction of aromatic ketones by a new ketoreductase from pichia glucozyma, Applied microbiology and biotechnology (2015) 1–9.
- [49] V. Prelog, Specification of the stereospecificity of some oxidoreductases by diamond lattice sections, Pure and Applied Chemistry 9 (1) (1964) 119–130.
- [50] P. Liang, B. Qin, M. Mu, X. Zhang, X. Jia, S. You, Prelog and anti-prelog stereoselectivity of two ketoreductases from candida glabrata, Biotechnology letters 35 (9) (2013) 1469 1473.
- [51] A. Li, L. Ye, F. Guo, X. Yang, H. Yu, Biocatalytic anti-prelog reduction of prochiral ketones with whole cells of a newly isolated strain empedobacter brevis zjuy-1401, Journal of Molecular Catalysis B: Enzymatic 117 (2015) 31–37.
- [52] R. Zhang, Y. Geng, Y. Xu, W. Zhang, S. Wang, R. Xiao, Carbonyl reductase scrii from candida parapsilosis catalyzes anti-prelog reaction to (s)-1-phenyl-1, 2-ethanediol with absolute stereochemical selectivity, Bioresource technology 102 (2) (2011) 483–489.
- [53] L. P. Tong, J. N. Cui, W.M. Ren, X. Y. Wang, X. H. Qian, Asymmetric bioreduction of substituted acenaphthenequinones using plant enzymatic systems: A novel strategy for the preparation of (+)-and (-)-mono hydroxyacenaphthenones, Chinese Chemical Letters 19 (10) (2008) 1179–1182.
- [54] The addition of water to the reaction is necessary for good agitation, because the water content in tomatoes is not enough.
- [55] In the case where the substrate is solid, we use a few quantity of ethanol just for solubilize it.
- [56] All prochiral ketones are reduced with correspondent procedure: NaBH4 in methanol at room temperature.
- [57] J. Seyden-Penne, H. C. Brown, Reductions by the alumino-and borohydrides in organic synthesis, John Wiley Sons, 1991.
- [58] S. Phothongkam, B.-J. Uang, Enantioselective reduction of ketones induced by a c2-symmetrical chiral hydroxyamide /titanium (iv) complex, Asian Journal of Organic Chemistry 4 (8) (2015) 794–799.
- [59] Tomatoes wastes are collected in Benamar Plant between June, 20 and July 15 in Wilaya of El- Tarf, Algeria.
- [60] For 100g of crude tomatoes 80% are the water and the rest is matter formed by different constituents.
- [61] T. Matsuda, T. Harada, N. Nakajima, K. Nakamura, Mechanism for improving stereoselectivity for asymmetric reduction using acetone powder of microorganism, Tetrahedron Letters 41 (21) (2000) 4135–4138.
- [62] A. A. Bothner-By, Simple preparation of optically active secondary alcohols1, Journal of the American Chemical Society 73 (2) (1951) 846–846.
- [63] Baker's Yeast and *Daucus Carota* for example.