

Research Journal of Pharmaceutical, Biological and Chemical

Sciences

The role of light microscope and classic culture in detection of periodontal pathogens, Review of the Literature

Kazem fatemi¹, Hamideh Sadat Mohammadipour², Ali Forouzanfar³

¹Oral & Maxillofacial Diseases Research Center, School of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran.

²Dental Materials Research Center, Department of Operative Dentistry, School of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran.

³Research Center for Patient Safety, Mashhad University of medical sciences, Mashhad, Iran.

ABSTRACT

More than 700 bacterial species are able to colonize the subgingival periodontal pocket of humans. Periodontitis is a multi-factorial chronic inflammatory disease of the periodontium with varying degrees of bone loss. Destruction of periodontium starts by colonization of gram-negative anaerobic microorganisms. Several methods have been used for identifying the involved species. The aim of this review is to identify the most virulent periodontal pathogens at the classic cultured medium.

Key words: Microscope, Classic culture, Porphyromonas gingivalis, Tannerella forsythia and Treponema denticola.



*Corresponding author Email id: Ali.forouzanfar@gmail.com.

March - April



INTRODUCTION

Although the human fetus is sterile, after birth several microorganisms start to colonize the whole body. After 2 years the entire human microbiota counts approximately 10¹⁴ microbial cells. On the other hand our body contains *10 times more bacteria* than human cells which comprises 2 kg of the total body and amazingly heavier than the brain's weight. The colonization of the oral cavity starts after birth by mainly facultative and aerobic bacteria. The first colonizers of the oral cavity include *Staphylococcus* spp, *Streptococcus* spp, *Lactobacillus* spp, *Actinomyces* spp, *Veillonella* spp and *Neisseria* spp. After tooth eruption, a more complex oral microbiota can be colonized at the teeth surfaces. Recent studies have revealed that about 500 different bacterial species have the ability to colonize the mouth and any individual may have around 50 to 150 different species [1, 2].

Discovery of dental plaque dates back to the seventeenth century, when Anton Von Leeuwenhoek, who invented the Microscope, saw microbial aggregates on the scraping materials of teeth surface [3, 4].

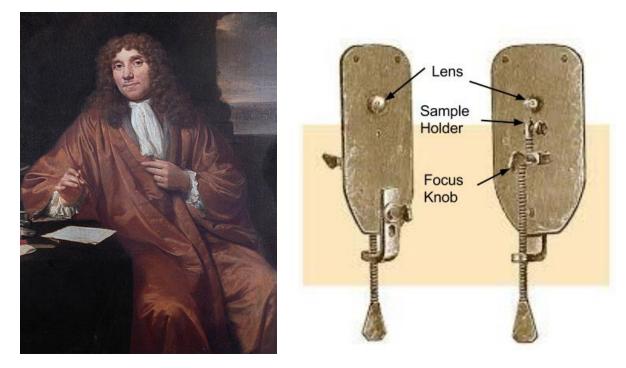


Fig 1. Antonie van Leeuwenhoek who invented the first microscopes for recognition of dental plaque bacteria and his microscope

Dental plaque is a structurally and functionally organized biofilm that form through several gradual stages. The first stage includes the formation of acquired dental pellicle, followed by the adhesion of first bacterial colonizers to the tooth surface. In this stage, the predominant teeth colonizers comprise mainly Actinomyces species and Streptococcus species, in particular Actinomyces israelii, Actinomyces naeslundii, Actinomyces oris, Streptococcus mitis, Streptococcus oralis, Streptococcus sanguinis, Streptococcus intermedius and Streptococcus gordonii. In the next stage secondary bacterial colonizers attach to the first colonizers using their specific surface molecules known as adhesins and receptors. This process is also called "Co-adhesion" or "Co-aggregation" which is necessary for the bacterial multiplication and synthesis of intermicrobial exopolymer matrix to form a mature biofilm. Secondary colonizers include mainly Campylobacter gracilis Campylobacter rectus, Campylobacter showae, Eubacterium nodatum, Aggregatibacter actinomycetemcomitans serotype b, Fusobacterium nucleatum spp nucleatum, Fusobacterium nucleatum spp vincentii, Fusobacterium nucleatum spp polymorphum, Fusobacterium periodonticum, Parvimonas micra, Prevotella intermedia, Prevotella loescheii, Prevotella nigrescens, Streptococcus constellatus, Tannerella forsythia, Porphyromonas gingivalis and Treponema denticola [5-9].

March – April

2016

RJPBCS

7(2)

Page No. 600



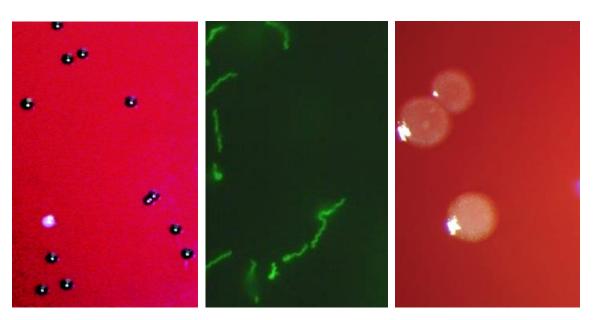


Fig 2. Red complex at the cultured medium. The left picture includes *Porphyromonas gingivalis microcolonies,* the middle *Tannerella forsythia and* the right picture *Spirochetes*.

Porphyromonas gingivalis

Porphyromonas gingivalis is an important periodontal pathogen that has been strongly investigated for its specific characteristics. This bacteris is a black-pigmented Gram-negative, anaerobic, non-motile, asaccharolytic rod with coccal to short rod appearance at the solid culture [Oliver & Wherry 1921] and has been intensely associated with severe forms of periodontal disease [10-13]. *Porphuromonas gingivalis* produce collagenase, several proteases for destroying immunoglobulins called "gingipain", hemolysins, hydrogen sulfide, fatty acids, endotoxin, ammonia, indole and etc [14-16].

Tannerella forsythia

Tannerella forsythia is a Gram-negative, anaerobic, spindle-shaped, highly pleomorphic rod that was first described in 1979 [17]. Growth of this species in the culture media is difficult and usually requires 7-14 days for small colonies to develop. *Tannerella forsythia* is associated more likely with gingivitis, chronic and aggressive periodontitis [18-21]. It has been reported that overweight or obese individuals have an overgrowth of *T. forsythia* compared to normal weight individuals and have a higher risk of developing periodontal disease [22]. *Tannerella forsythia* requires N-acetyl muramic acid for the growth which can be provided by co-cultivation with F. *nucleaium* in the medium culture [23]. Virulence determinants of this species include Surface associated glycoproteins (S-layer, TfsA and TfsB) for attachment to the epithelial cells, BspA surface protein for adherence and invasion and interaction with the innate host response via TLR₂ and TLR₃, Sialidases (SiaHI) for degradation of host oligosaccharides and Protease (PrtH) for epithelial barrier disturbance [24-26].

Spirochetes

Spirochetes are Gram-negative, anaerobic and highly motile bacteria that are visualized as helicalshaped colonies in the strict anaerobic conditions and a specific medium culture. Several studies have demonstrated the role of this microorganism in the etiology of destructive periodontal diseases. The most important subspecies of this group includes *Treponema denticola*, *Treponema vinccentii and Treponema Socranski* and *Treponema pallidum*. The numbers of this group interestingly increase in sites with increased pocket depth. Of this species *Treponema denticola* has gained more interest because of membership in red complex which is the most destructive periodontal pathogenic complex [27-29] and having several virulent factors including: Major sheath protein (Msp) which is a cell surface protein associated with adherence, Leucine rich protein (Lrr) for bacterial and epithelial cell adherence, Dentilisin (PrtP) for degradation of host cell matrix proteins and Trypsin-like protease (OpdB) for Protein and peptide degradation [30, 31].

March – April

2016

RJPBCS

7(2)

Page No. 601



CONCLUSION

Periodontitis is a prevalent and progressive disease that affects individuals with poor oral hygiene and lead to the loosening and finally loss of teeth. Other etiologic factors may include environmental conditions, genetic polymorphism, nutritional status and the psychological stresses that can affect disease development and progression. The destruction of periodontitis is believed to result from a mixed bacterial infection of the periodontal structure, and several clinical studies have demonstraded the role of *Porphyromonas gingivalis, Tannerella forsythia and Treponema denticola* in the pathogenesis of the disease. For this reason identifying specific pathogens via laboratory experimental approaches of the cultured medium and several physical or chemical anti-plaque strategies have been applied [32-37].

ACKNOWLEDGEMENT

I take this opportunity to appreciate all of the Mashhad Department of Periodontology and Implant dentistry faculty members for their help and support. The alphabetical list of professors includes Hamid Reza Arab, Seyed Ali Banihashemrad, Kazem Fatemi, Habib allah Ghanbari, Amir Moien Taghavi, Majid Reza Mokhtari, Mehrdad Radvar, Mohammad Ebrahim Rahmani, Naser Sargolzaei and Mahmoud Tamizi. I thank all of them for the constant encouragement, support and attention.

Declarations of Interests

The authors confirm that this article content has no conflict of interest.

REFERENCES

- Quirynen M, Teughels W, Haake SK, Newman MG. Microbiology of Periodontal Diseases. In: Newman MG, Takei HH, Klokkevold PR, Carranza FA, editors. Carranza's Clinical Periodontology. 10th ed. St Louis, Missouri: Elsevier (Saunders); 2006. pp. 134–69.
- [2] Lang NP, Mombelli A, Attstrom R. Oral Biofilms and Calculus. In: Lindhe J, Lang NP, Karring T, editors. Clinical Periodontology and Implant Dentistry. 5th ed. Oxford: Blackswell- Munksgaard; 2008. pp. 183–267.
- [3] Socransky SS, Haffajee AD. Dental biofilms: difficult therapeutic targets. Periodontol. 2000; 2008(28):12–55.
- [4] Rita CH, Priyank B, Ruchi B. Biofilms: A microbial home. J Indian Soc Periodontol. 2011 Apr-Jun; 15(2): 111–114.
- [5] Socransky, S.S. & Haffajee, A.D. (2002). Dental biofilms: difficult therapeutic targets. *Periodontology* 2000 28, 12–55.
- [6] Marsh, P.D. & Bowden, G.H.W. (2000). Microbial community interactions in biofilms. In: Allison, D.G.,
 Gilbert, P, Lappin- Scott, H.M. & Wilson, M., eds. *Community Structure and Co-operation in Biofilms*.
 Society for Microbiology Symposium 59. Cambridge: Cambridge University Press, pp. 167–198.
- [7] Kolenbrander, P.E., Palmer, R.J., Jr., Rickard, A.H. *et al.* (2006). Bacterial interactions and successions during plaque development. *Periodontology 2000* 42, 47–79.
- [8] Marsh, P.D. & Devine, D.A. (2011). How is the development of dental biofilms influenced by the host? Journal of Clinical Periodontology 38 Suppl 11, 28–35.
- [9] Kolenbrander, P.E., Palmer, R.J., Jr., Periasamy, S. & Jakubovics, N.S. (2010). Oral multispecies biofilm development and the key role of cell-cell distance. *Nature Reviews Microbiology* **8**, 471–480.
- [10] Melvin W L, Assad D A, Miller G A, Gher M E, Simonson L, York A K. Comparison of DNA probe and ELISA microbial analysis methods and their association with adult periodontitis. J Periodontol.1994; 65:576–582.
- [11] Preus H R, Anerud A, Boysen H, Dunford R G, Zambon J J, Loe H. The natural history of periodontal disease. The correlation of selected microbiological parameters with disease severity in Sri Lankan tea workers. J Clin Periodontol. 1995; 22:674–678.
- [12] Socransky S S, Haffajee A D, Smith C, Dibart S. Relation of counts of microbial species to clinical status at the sampled site. J Clin Periodontol. 1991; 18:766–775.
- [13] Wolff L F, Aeppli D M, Pihlstrom B, Anderson L, Stoltenberg J, Osborn J, Hardie N, Shelburne C, Fischer
 G. Natural distribution of 5 bacteria associated with periodontal disease. J Clin Periodontol.1993; 20:699–706.

7(2)



- [14] Darveau, RP. Belton, C.M., Reife, RA & Lamont, RJ. (1998). Local chemokine paralysis, a novel pathogenic mechanism for *Porphuromonas gingioalis*. *Infection and Immunity 66*, 1660-1665.
- [15] Fletcher, J., Nair, S., Poole, S., Henderson, B. & Wilson, M. (1998). Cytokine degradation by biofilms of *Porphuromonas gingivalis. Current Microbiology* 36, 216-219.
- [16] Sandros, LKarlsson, C, Lappin, D.F., Madianos. P.N., Kinane, D.P. & Papapanou, P.N. (2000).Cytokine responses of oral epithelial cells to *Porphyromonas gingivalis*. *Journal of Dental Research* 79, 1808-1814.
- [17] Tanner, ACR, Haffer, C, Bratthall, G.T., Visconti, RA & Socransky, S.s. (1979). A study of the bacteria associated with advancing periodontitis in man. *Journal of Clinical Periodontology* 6, 278-307.
- [18] Dzink JL, Socransky SS, Haffajee AD. The predominant cultivable microbiota of active and inactive lesions of destructive periodontal diseases. J Clin Periodontol. 1988; 15:316–323.
- [19] Grossi SG, Genco RJ, Machtei EE, Ho AW, Koch G, Dunford RG, Zambon J, Hausmann E. Assessment of risk for periodontal disease. II. Risk indicators for alveolar bone loss. J Periodontol.1995; 66:23–29.
- [20] Listgarten MA, Lai CH, Young V. Microbial composition and pattern of antibiotic resistance in subgingival microbial samples from patients with refractory periodontitis. J Periodontol. 1993;64:155– 161.
- [21] Tanner A, Maiden MF, Macuch PJ, Murray LL, Kent RL., Jr Microbiota of health, gingivitis, and initial periodontitis. J Clin Periodontol. 1998; 25:85–98.
- [22] Haffajee AD, Socransky SS. Relation of body mass index, periodontitis and *Tannerella forsythia*. Journal of Clinical Periodontology. 2009; 36:89–99.
- [23] Wyss, C. (1989). Dependence of proliferation of *Bacteroides forsythus* on exogenous Nacetylmuramic acid. *Infection & Immunity* 57, 1757-1759.
- [24] Lee, S.W., Sabet, M., Um, H.S. et al. (2006). Identification and characterization of the genes encoding a unique surface (S-) layer of Tannerella forsythia. Gene 371, 102–111.
- [25] Sharma, A., Sojar, H.T., Glurich, I. *et al.* (1998). Cloning, expression, and sequencing of a cell surface antigen containing a Leucine rich repeat motif from *Bacteroides forsythus* ATCC 43037. *Infection and Immunity* **66**, 5703–5710.
- [26] Saito, T., Ishihara, K., Kato, T., Okuda, K. (1997). Cloning, expression, and sequencing of a protease gene from *Bacteroides forsythus* ATCC 43037 in *Escherichia coli*. *Infection and Immunity* **65**, 4888–4891.
- [27] Chan EC, McLaughlin R. Taxonomy and virulence of oral spirochetes. *Oral Microbiol Immunol. 2000 Feb; 15(1):1-9.*
- [28] Choi BK, Paster BJ, Dewhirst FE, Gobel UB. (1994). Diversity of cultivable and uncultivable oral spirochetes from a patient with severe destructive periodontitis. Infect Immun 62:1889-1895
- [29] Holt SC, Ebersole JL. (2005). *Porphyromonas gingivalis, Treponema denticola*, and *Tannerella forsythia*: the "red complex", a prototype polybacterial pathogenic consortium in periodontitis. Periodontol 200038:72-122.
- [30] Fenno, J.C. & McBride, B.C. (1998). Virulence factors of oral treponemes. Anaerobe 4, 1–17.
- [31] Rosen, G., Genzler, T. & Sela, M.N. (2008). Coaggregation of *Treponema denticola* with *Porphyromonas gingivalis* and *Fusobacterium nucleatum* is mediated by the major outer sheath protein of *Treponema denticola*. *FEMS Microbiology Letters* 289, 59–66.
- [32] Forouzanfar A, Clinical application of antibiotics for the management of periodontal disease, a systematic review, American Journal of Pharmaceutical Sciences, Volume 1, Issue 1, 2015, 39-44.
- [33] Forouzanfar A, The role of mouth rinses in treatment of oral and periodontal disease, a systematic review article, Australian Journal of Pharmaceutical Research, Volume 2, Issue 1, 2015, 1-5.
- [34] Arab H, Maroofian A, Golestani S, Shafaee H, Sohrabi K, Forouzanfar A. (2011). Review of the therapeutic effects of Camellia sinensis (green tea) on oral and periodontal health. Journal of Medicinal Plants Research, 5(23), 5465-5469.
- [35] M. Amirchaghmaghi, J. Salehinejad, M. Basirat, Z. Delavarian, A. Javadzade and A. Forouzanfar, 2010. Gingival Ancient Schwannoma: Review of Literature and a Case Report. Journal of Applied Sciences, 10: 3137-3140.
- [36] Kazem Fatemi, Hamideh Sadat Mohammadipour and Ali Forouzanfar, 2016. Alopecia Areata Associated with Generalized Mild Chronic Periodontitis: A Case Report and Review of the Literature. Research Journal of Pharmaceutical, Biological and Chemical Sciences, 7(1), 1563-1566.
- [37] Ghanbari H, Mousavi SA, Forouzanfar A, Zakeri M, Shafaee H, Shahnaseri S. Synergic phototoxic effect of visible light or Gallium-Arsenide laser in the presence of different photosensitizers on *Porphyromonas gingivalis* and *Fusobacterium nucleatum*. Dent Res J 2015; 12:323-30.

7(2)