



Research Journal of Pharmaceutical, Biological and Chemical Sciences

Phytochemical Analyses of the Methanol, Hot Water and N-Hexane Extracts of the Aerial Parts of *Cassytha Filiformis* (Linn) and Leaves of *Cleistopholis Patens* (Benth)

Adonu Cyril C^{*1}, UGWU Okechukwu PC², ESIMONE CO¹, OSSAI Emmanuel C², BAWA
Abubakar³, NWAKA Andrew C⁴ and OKORIE Chris U⁴

1Department of Pharmaceutics, University of Nigeria, Nsukka, Enugu State, Nigeria.

2Department of Biochemistry, University of Nigeria, Nsukka, Enugu State, Nigeria.

3Department of Preliminary and Remedial Studies, Federal Polytechnic Mubi, Adamawa State, Nigeria.

4Department of Biochemistry, Anambra State University Uli, Anambra State, Nigeria.

ABSTRACT

Phytochemical studies of the aerial parts of *Cassytha Filiformis* Linn and leaves of *Cleistopholis patens* Benth were analysed. The phytochemical analyses of the methanol, hot water and n-hexane extracts of *Cassytha Filiformis* Linn and *Cleistopholis patens* showed the presence of glycosides, steroids, terpenoids, alkaloids, carbohydrates, acidic compounds, resins, saponins, tannins, fats and oil and flavonoids. Phytochemical like proteins was not detected.

Keywords: *Cassytha Filiformis* Linn, *Cleistopholis patens* Benth, Phytochemicals, Flavonoids and acidic compounds.

**Corresponding author*

INTRODUCTION

People on all continents have used hundreds to thousands of indigenous plants for the treatment of ailments since pre-historical time. Spices and herbs are generally known to possess antibacterial and antioxidant properties [20]. Many medicinal plants are used in modern medicine where they occupy a very significant place as raw material for important drugs and plants [12]. Medicinal plants are of great importance to the health of individual and communities. Phytochemicals are chemical compounds that are naturally found in plants. They are responsible for the colour and organoleptic properties of the plant. Phytochemicals could be available as dietary supplements, but the potential health benefits of phytochemicals are derived from consumption of the whole plant [13].

Cassytha filiformis is a leafless, perennial, climbing, twinning, vine-like, autoparasitic and plant-hyperparasitic phanerogram (seed-bearing plant) in the plant family *Lauraceae*. The stems, which are green to orange, filiform and glabrous are long and thread-like, measuring 3-8m in length, grows in a triangle on hosts. Flowers are sessile and few in spicate inflorescence 1-2cm long. *Cassytha filiformis* Linn inhabits coastal vegetation, usually on sandy dunes, margins of evergreen gully and venerin forests, deciduous and coastal woodlands, montane grassland and dambo margins and vegetation of sandy beaches or lakes. It grows on small bushes, woods shrubs and low tree, grasses and herbaceous weeds [15]. Its seeds might be dispersed by animals, man, water, strong winds, farm machinery. They parasitize other plants by specialized attachment and penetration with feeding structures known as *haustoria*. According to [16], "this plant is distributed throughout India and is used for medicinal purposes in China, Madagascar and South Africa.

In medicine, it is used traditionally for treatment of some human birthing issues. Modern midwives recommend taking the juice made from crushed vines for 4 weeks before the expected date of birth in order to ease labour pains and to quicken labour time and lubricate the birth canal [9]. *Cassytha filiformis* is also purported to be used by several different Polynesian cultures for the treatment of cancers. In modern medical research, *Cassytha filiformis* has a number of biologically active chemical compounds with potential human health application. Ocoteine isolated from *Cassytha filiformis*, is an alpha-adrenoreceptor antagonist in rat thoracic aorta, have antiplatelet aggregation activity [1]. *Cassytha filiformis* is used as vasorelaxant [20] and adrenoreceptor antagonist [1], antitrypanosomal agent [1] and diuretics in traditional medical practice [8].

Cleistopholis patens (Benth) is a tree 20-30m tall, with a trunk up to 10m tall and 20-90cm wide, its bark is grayish-white, smooth fibrous or furrowed. It is a fast growing, commonly seen in forests and rapidly colonizing abandoned areas. The bole is slender, cylindrical and straight and its timber is straight grained. The tree is sun-loving, common in distributed forest and rapidly colonizing abandoned areas. It is used in traditional medical practices in many parts of Africa where it has several applications. [6] isolated oligorhamnosides anti-bacteria from *Cleistopholis patens* Benth. In addition, alkaloids, (eupolauridine) and 3-methoxy champagine, from the ethanolic extract of *Cleistopholis patens*

Benth are potent anti-fungal agents [10] reported that the steroidal, glycoside and alkaloidal fractions of *Cleistopholis patens Benth* are effective against *klebsiella pneumonia*.

This study was designed to determine the phytochemical constituents of hot water, methanol and n-hexane extracts of the aerial parts of *Cassytha Filiformis Linn* and leaves of *Cleistopholis patens Benth*.

MATERIALS AND METHODS

Plant material

Fresh aerial parts of *Cassytha Filiformis Linn* and leaves of *Cleistopholis patens Benth* were collected from Nsukka in Enugu State Nigeria between June to August, 2009. The aerial parts and leaves were indentified by Mr. A.O Ozioko of the Bioresource development and conservation programme (BDCCP) Nsukka, Enugu State, Nigeria.

The two plants were dried under the shade for 10 days, pulverized and stored at room temperature for two weeks before extraction processes.

Chemicals/Reagents

All chemicals used in this study were of analytical grade and products of May and Baker, England; BDH, England and Merck, Darmstand, Germany.

Extraction Procedure

The fresh aerial parts of *Cassytha Filiformis Linn* and leaves of *Cleistopholis patens Benth* plant were plucked and shade dried for 10 days, after which the two plants were pulverized into coarse form with a crestor high speed milling machine and stored for two weeks. Approximately, 400g each of the coarse forms of *Cleistopholis patens Benth* and *Cassytha Filiformis Linn* were then macerated in 2.0L and 4.0L of hot water respectively and allowed to stand at room temperature. This was left to stand for 48 hours. After that the hot water extracts were filtered through muslin cloth on a plug of glass wool in a glass column. The resulting hot water extracts were concentrated and evaporated to dryness using rotary evaporator at an optimum temperature of between 40 and 45°C to avoid denaturation of the active ingredients. The concentrated extracts were then stored in the refrigerator. The methanol and n-hexane extracts of the two plants were obtained by soxhlet extraction. Approximately, 300g of pulverized *Cassytha Filiformis Linn* and 400g of pulverized leaves of *Cleistopholis patens Benth* were extracted with 1.5L and 1.6L of menthanol respectively. Finally, 400g of pulverized *Cassytha Filiformis Linn* was extracted with 1.5L n-hexane.

Determination of yield of extracts

The percentage yield of the extracts were determined by weighing the coarse *Cassythia Filiformis Linn* and *Cleistopholis patens Benth* before extraction and *Cassythia Filiformis Linn* and leaves of *Cleistopholis patens Benth* extracts after concentration and then calculated using the formula.

$$\text{Percentage (\%) yield} = \frac{\text{Weight (g) of the concentrated extracts}}{\text{Weight (g) of the ground plant extracts}} \times 100$$

Determination of phytochemicals

The qualitative phytochemical analyses of the crude hot water, methanol and n-hexane extracts of both plants' parts were carried out based on the procedure outlined [5] and [19].

RESULTS

Percentage yield of the extracts

The percentage yield of all extracts of *Cleistopholis patens Benth* were greater than that for *Cassythia Filiformis Linn*. The yield per plant extract increased with increasing polarity of the solvent; highest yield was noted with hot water, followed by methanol and the least was n-hexane extract.

Table 1: Percentage yield of the extracts of aerial parts of *Cassythia Filiformis Linn* and *Cleistopholis patens Benth*

Plant	Part	Percentage yield of the extracts (%)		
		Hot water	Methanol	n-hexane
<i>Cassythia Filiformis Linn.</i>	Aerial parts	12.5	5.8	12
<i>Cleistopholis patens Benth</i>	Leaves	12.9	10.5	-

The results from Table 2: shows the presence of glycosides, steroids, terpenoids, alkaloids, carbohydrates, acidic compounds, resins, saponins, tannins, fats and oil and flavonoids. Phytochemical like proteins was not detected.

Table 2: Phytochemical composition of the extracts of aerial parts of *Cassytha Filiformis* Linn and *Cleistopholis patens* Benth .

Phytoconstituents	Cassytha Filiformis Linn			Cleistopholis patens Benth		
	MF	AF	NF	MP	AP	NP
Glycosides	+++	++++	-	++	+	-
Steroids	-	-	++	+	-	+
Terpenoids	+	-	++	+++	-	-
Alkaloids	+	+++	-	+	-	-
Acidic compounds	-	+	-	-	-	-
Carbohydrates	+	++	-	++	+	-
Resins	-	-	+++	-	+	++
Tannins	++	+++	-	-	+	-
Saponins	-	-	-	+++	+	-
Flavonoids	+	-	++	++	-	-
Proteins	-	-	-	-	-	-
Fat and oils	-	-	+	++	-	+

Key to abbreviations

MF: Methanol extracts of *Cassytha Filiformis* Linn aerial parts

AF: Hot water extracts of *Cassytha Filiformis* Linn aerial parts

NF: n-Hexane extracts of *Cassytha Filiformis* Linn aerial parts

MP: Methanol extracts of *Cleistopholis patens* Benth leaf

AP: Hot water extracts of *Cleistopholis patens* Benth leaf

NP: n-Hexane extracts of *Cleistopholis patens* Benth leaf

++++ : Abundantly present

+++ : Copiously present

++ : Moderately present

+ : Slightly present

- : Absent

DISCUSSION

The phytochemical analysis of the aerial parts of *Cassytha Filiformis* Linn revealed the presence of flavonoids, alkaloids, glycosides, terpenoids, steroids, fats and oil, carbohydrates, resins and tannins, but proteins and saponins were found to be absent in the extract. In the methanol extracts of the same plants glycosides were

Copiously present whereas other constituents occurred in moderate or slight amounts. While, in hot water extracts, glycosides occurred at abundant level, alkaloids and tannins at copious amounts and carbohydrates at low level. Steroids, terpenoids, flavonoids and reducing sugars exist at substantial amounts in n-hexane fraction of the plant. These metabolites have been shown to be responsible for various therapeutic activities of medicinal plants [19]. For instance, flavonoids acts as a “biological response modifiers” such as anti-allergic, anti-inflammatory, ant-microbial [3], and anti-cancer activities shown from *in vitro* studies [11], [14]. According to [2] flavonoids are especially known for their anti-fungal effects against wide array

of micro-organisms, the activities are attributed to their abilities to complex with extracellular and soluble proteins with microbial cell wall.

Tannins have been found to form irreversible complexes with proline proteins resulting in inhibition of cell protein synthesis. Tannins are potent antioxidants and also used for treating diarrhea and dysentery. Alkaloids, the largest group of chemicals, produced by plants have many biological activities. Alkaloids (pure and synthetic) isolates are used as basic medicinal agents for their analgesic, antispasmodic, bactericidal effects [17]; [11] and antifungal effects [10]. More so, saponins have been found to intercalate DNA of microorganism as mechanism of their antimicrobial activity [7] and thus exhibit antifungal activity [2].

The phytochemical test of the second plant parts, the leaf extracts of *Cleistopholis patens Benth* also revealed the presence of glycosides, steroids, terpenoids, alkaloids, saponins, flavonoids and carbohydrates. Acidic compound and proteins were not detected in all the extracts. The methanol leaf extract of this plant showed the presence of copious amount of terpenoids and saponins, with substantial amount of glycosides, steroids, alkaloids and saponins in the same extract. Hot water and n-hexane extracts of this plant parts revealed the absence of most important secondary metabolites known to have antimicrobial activity [3], [2], [17] and [11]. Even where these important metabolites occur, they exist at low level. The absence of major metabolites in the hot water and n-hexane leaf extracts of *Cleistopholis patens Bent* might be the reason for their negligible antimicrobial activities against the test microorganisms.

Futher evaluation, isolation and characterization of these metabolites are therefore encouraged in order to elucidate the active agents responsible for the observed numerous medicinal effects of these plants.

REFERENCES

- [1] Chang CW, Ko FN, Su MJ, et al. Japanese J Pharm 1997; 73: 202-214.
- [2] Cowan MW. Clin Microb Rev 1999; 12: 564-582.
- [3] Cushnie TPT and Lamb AJ. Int J Antimicrob Agents 2005; 26: (5):343-356.
- [4] Elvin-Lewis M. J Ethnopharmacol 2001; 75: 141-164.
- [5] Harborne J. Phytochemical Methods. A Guide to Modern Technique of Plant Analysis. Chapman and Hall. Thompson Science, London. pp.107.
- [6] Hu JF, Garo E, Hough GW, et al. J Nat Prod 2006; 69 (4): 585-590.
- [7] Jennins BR and Ridder PJ. Biophys Stru Mech 1983; 10: 71-79.
- [8] Kirtikar KR and Basu BD. Indian Medicinal plants. 2nd edition. 1991; pp. 1741-2163.
- [9] Kobayashi J. J Trop Paediatr 1976; 22: 260-262.
- [10] Liu S, Oguntimein B, Hufford CD, et al. Antimicrob Agents Chemother 1990; 8: 529-593.
- [11] Okwu DE. J Sustain Agri Environment 2004; 6(2): 140-147.
- [12] Rajendra CE, Gopal SM, Mahaboob AN, et al. International Journal of Pharmacognosy and Phytochemical Research 2011; 3(3): 61-63.
- [13] Rao AV and Rao LG. Pharmacol Res 2007; 55: 16-207.



- [14] Salah N, Niller NJ, Pangange G, et al. Biochem Broph 1995; 2: 339-346.
- [15] Scot CN. Plant disease PD-42: http://www.ctahr.hawaii.edu/free_pubs.s20/8/2010
- [16] Sharm S, Hullatti Kk, Prasanna SM, et al. Pharma Res 2009; 1: 30-327.
- [17] Stary F. The National Guide to Medicinal Herbs and Plants. Tiger books International London 1998; pp 12-16.
- [18] Talalay P and Talalay P. Academic Medicine 2001; 76: 3-238.
- [19] Trease GE and Evans WC. Pharmacognosy 13th Edition. ELBS Oxford University Press, London, UK 1989; pp. 245-263.
- [20] Wu Yang-Chang, Chang F-R, Ya-Chief C et al. Phytoche Res 1998; 12: 539-541.