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The Composition And Role Of The Microbiota Of Chickens' Gastrointestinal Tract.

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ABSTRACT

This article reflects current data on the study of the ecosystem of microorganisms inhabiting the poultry digestive tract. Information on the use of T-RFLP analysis (Terminal restriction fragment length polymorphism) - a molecular genetic method based on an analysis of the length polymorphism of amplified restriction DNA fragments of microorganisms in the study of the taxonomic diversity of microbial populations of the gastrointestinal tract and results obtained using this method is presented. The fact that about 800 species of microorganisms from nearly 300 genera have now been found in the poultry intestine, and less than 10% of this biodiversity. Moreover, many species and even genera of intestinal microorganisms detected by metagenomic methods are still not identified.

Keywords: microbiota of the gastrointestinal tract of birds, T-RFLP analysis.



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SHORT REVIEW

The problem of studying the ecosystem of microorganisms inhabiting the poultry digestive tract is highly relevant. The important role of the microorganisms of the gastrointestinal tract (GIT) in chickens is generally recognized: they provide the host organism with certain nutrients through the use of its own enzymes (amylases, cellulases and others), as well as antibiotic substances, proteins, hormones, vitamins and a number of other compounds [19, 20, 21]. Most information about the microflora inhabiting the gut of chickens, obtained using classical methods of microbiology, according to which the basis of the microbial community are bifidobacteria, streptococci, lactobacilli, lactate-fermenting bacteria, eubacteria, bacteroids and entero bacteria [12, 20, 21].

Unique molecular genetic methods open up to a researcher studying the composition of the intestinal microbiome of chickens, which demonstrated that it contains representatives of up to 140 genera of bacteria, of which only 10% are identified by the 16S rRNA gene, and the rest belong to new species or even new genera [1, 2, 4].

One of the most promising today is the T-RFLP analysis (Terminal restriction fragment length polymorphism) - a molecular genetic method based on an analysis of the length polymorphism of amplified restriction DNA fragments of microorganisms [22]. It is designed to determine the number, relative abundance and taxonomic affiliation of all bacteria in the microbial ecosystem. This allows wide and deep comparative study of microbiological communities in their development and change.

For the first time in Russia, based on an analysis of foreign studies, the T-RFLP methodology was modified to study the microflora of the gastrointestinal tract of agricultural birds [13, 14]. Recent studies of the intestine have allowed us to suggest that the gastrointestinal tract of a bird (for example, broilers) contains about 640 species of bacterial microorganisms. The number and variety of intestinal microflora vary in different parts of the intestine, and it is obvious that the departments with less favorable conditions (stomach) or faster passage of the contents of the intestine (duodenum) have fewer bacteria [12].

Modern molecular genetic analysis methods of the microbiome - real-time PCR, NGS, T-RFLP - have provided new insights into the real taxonomic diversity of the microbial populations of the gastrointestinal tract [1, 9, 12]. About 800 species of microorganisms from almost 300 genera have been found in the poultry intestine [9], and only less than 10% of this biodiversity can be isolated and cultured by laboratory [7]. Moreover, many species and even genera of intestinal microorganisms detected by metagenomic methods are still not identified [8].

It was previously believed that the digestive canal of a newly hatched chicken is microbiologically sterile and that it is inoculated with microorganisms in the first hours and days after hatching [21]. However, in recent studies, intestinal microflora was found even in 6-day-old chicken embryos, although the taxonomic diversity of embryonic flora is much less than in chickens a few days after hatching [2, 16, 17, 22].

Modern studies carried out using the modern molecular genetic method T-RFLP allowed us to establish quite clear standards for the maintenance of some major groups of microorganisms in the blind processes of the gastrointestinal tract of healthy broiler chickens and allowed us to conclude that the assumption that was based on the data of classical microbiological experiments that the main part of the microflora of the blind processes of the gastrointestinal tract of birds are bifidobacteria, lactobacilli and bacteroids[21] is incorrect.

Normal microflora is symbiotic with respect to the host organism, performing a number of important functions that are not limited to digestion. It participates in the function of the cardiovascular, endocrine, hematopoietic, nervous, immune and other systems of the host organism; produces a number of useful metabolites for it, such as endogenous antibiotics, proteins, enzymes, hormones, vitamins, low molecular weight fatty acids, etc.; serves as a barrier against invasion and uncontrolled reproduction of pathogenic microorganisms; participates in the neutralization of toxic xenobiotics; modulates gene expression in the intestines [5, 6, 10, 15, 18, 23, 24.



It has been established that bacteriocins produced by probiotic strains *L. salivarius* and *Paenibacilluspolymyxa* function as colonizing peptides, facilitating the introduction and / or domination of the producer in a niche already occupied by him [26]. These probiotic strains showed pronounced antagonistic activity against *C. jejuni* in vitro, but were inactive in vivo [26]. Based on the data obtained, it was hypothesized that the primary structure of the bacteriocin synthesized in vivo under the influence of various internal factors may differ from the primary structure of the peptide synthesized in vitro.

Along with mobility and adhesion, the invasion is an important step in the pathogenesis of campylobacteriosis. Probiotics are able to reduce invasion due to their competitive properties. Of considerable interest study in which the coaggregation of lactobacilli with *C. jejuni* is studied [27]. In evolution, bacterial coaggregation emerged as an integral process in the development of multi vid biofilms [28]. An example of a multi-species biofilm is biofilm formed on human teeth [29]. However, some microorganisms, in particular, lactobacilli, use co-aggregation with *C. jejuni* to neutralize virulence factors and subsequent competing killing to ensure colonization resistance of the microecological niche of the host [27].

CONCLUSION

Thus, the analysis of current data on the study of the taxonomic diversity of the microbial populations of the bird's gastrointestinal tract suggests that the taxonomic diversity is much greater than that revealed by classical methods of microbiology, which opens up broad horizons for studying this issue.

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REFERENCES

- [1] Amit-Romach E., Sklan D., Uni Z. Microflora ecology of the chicken intestine using 16S ribosomal DNA primers/E. Amit-Romach. Poult. Sci. 2004. 83(7). pp. 1093-1098.
- [2] Apajalahti J., Kettunen A., GrahamH. Characteristics of the gastrointestinal microbial communities, with special reference to the chicken. World's Poult. Sci. J. 2004. 60. pp. 223-232.
- [3] Kizewetter-Swid M., Binek M. Bacterial microflora of the chicken embryos and newly hathed chicken.J. Anim. Feed Sci. 2008. 17. pp. 224-232.
- [4] Manukyan V.A., Laptev G.Yu., Nikonov I.N., Ignatova G.V., Aidinyan G.T. Cellobacterin-T in broiler diets. XIVth European Poultry Conference Information and Proceedings. Birger Svihus, Editor. 2014. P. 559.
- [5] Rantala M., Nurmi E. Prevention of the growth of Salmonella infantis in chickens by flora of the alimentary tract of chickens. Br. Poult. Sci. 1973. 14, 6. pp. 627-630.
- [6] Rigby C.,PettitJ. Some factors affecting Salmonella typhymurium infection and shedding in chickens raised on litter.Avian Dis. 1979. 23. pp. 442-455.
- [7] Torok V.A., Ophel-Keller K., LooM. Application of methods for identifying broiler chicken gut bacterial species linked with increased energy metabolism. Appl. Environ. Microbiol. 2008. 74(3) pp. 783-791.
- [8] Torok V.A., Hughes R.J., Mikkelsen L.L. *et al*.Identification and characterization of potential performance-related gut microbiota in broiler chickens across various feeding trials. Appl. Environ. Microbiol. 2011. 77(17). pp. 5868-5878.
- [9] Xiang Zhu Y., Zhong T., Pandya Y. *et al.*16SRNA- based analysis of microbiota from the cecum of broiler chickens. Appl. Environ. Microbiol. 2002. 68(1). pp.124-137.
- [10] Yin Y.,LeiF., ZhuL., LiS., WuZ. *et al*. Exposure of different bacterial inocula to newborn chicken affects gut microbiota development and ileum gene expression. ISMEJ. 2010. 4. pp. 367-376.
- [11] Burmistrov V.A. Normal microflora and its importance for human health. Drugs for the prevention and treatment of dysbacteriosis. Novosibirsk: Vector-Vita, 2009. P. 19.
- [12] Grozina A.A. The composition of the microflora of the gastrointestinal tract in broiler chickens when exposed to probiotic and antibiotic (according to T-RFLP RT-PCR). Agricultural Biology. 2014. 6. pp. 46-58.
- [13] Egorov I., Laptev G., Nikonov I., Kryazhevskikh L. T-RFLP analysis of intestinal microflora the basis of the choice of feed additives for poultry. Poultry farming. 2010. 9. P. 25.



- [14] Egorova T.A., Lenkova T.N., Il'ina L.A. et al.Effect of probiotics based on Saccharomyces sp. and Bacillus subtilis on the bacterial community of the blind processes of the intestines and the productivity of broiler chickens. Agricultural Biology. 2016. 51(6). pp. 891-902.
- [15] Zon G. Autochthonous bird microflora. Poultry farming. 1992. 6. pp. 20-22.
- [16] Ilina L.A., Yıldırım E.A., Nikonov I.N., Filippova V.A., Yu G. Laptev et al. Metagenomic landscape of the gastrointestinal tract of chick embryos using the T-RFLP (L.A.A.) method. Reports of RAS. 2016. 466(4). P. 482.
- [17] Ilina L.A., Yıldırım E.A., Novikova N.I., Nikonov I.N., Filippova V.A. et al. Microflora in ovo: the possibilities of the molecular-biological method T-RFLP. Bulletin of Russian agricultural science. 2015.
 6. pp.6-8.
- [18] Pavlova N., Kirzhayev F., LaCite R. The value of normal microflora of the digestive tract of birds for their bodies. BIO. 2002. 1. pp. 4-7.
- [19] Podobed L.I., Laptev G.Yu., Kapitonov E.A., Nikonov I.N. Optimization of digestion and protein nutrition of poultry.SPb: Right PRINT SOUTH, 2017. P. 348.
- [20] Tarakanov B.V. Methods for studying the microflora of the digestive tract of farm animals and poultry. B.V. Cockroaches. M., 2006. P. 128.
- [21] Timoshko M.A. The microflora of the digestive tract of farm animals. Chisinau: Stinnitsa, 1990. P. 161.
- [22] Fisinin V.I., Laptev G.Yu., Nikonov I.N., Ilina L.A., Yıldırım E.A. et al Changes in the bacterial community in the gastrointestinal tract of chickens during ontogenesis. Agricultural Biology. 2016. 51(6). pp. 883-890.
- [23] Chakhava O.V., Gorskaya E.M., Ruban S.Z. Microbiological and immunological bases of gnotobiology.M.: Medicine, 1982. P. 159.
- [24] Shenderov B.A. Medical microbial ecology and functional nutrition. Vol.1: Microflora of humans and animals and its functions. M .: "Grant", 1998. P. 288.
- [25] Fisinin V.I., Laptev G.Yu., Egorov I.A. et al.Modern ideas about the intestinal microflora of poultry in different diets: molecular genetic approaches. SergievPosad, 2017. P. 263.
- [26] Stern N.J., Eruslanov B.V., Pokhilenko V.D., Kovalev Y.N., Volodina L.L., Perelygin V.V., Mitsevich E.V., Mitsevich I.P., Borzenkov V.N., Levchuk V.P., Svetoch O.E., Stepanshin Y.G., Svetoch E.A. Bacteriocins reduce Campylobacter jejuni colonization while bacteria producing bacteriocins are ineffective. MicrobEcol Health Dis. 2008.20(2). pp. 74-79.
- [27] Romero M., Coaggregation of Lactobacilli with Campylobacterjejuni. I-ACES 2014. 1.
- [28] Rickard A.H., Gilbert P., High N.J., Kolenbrander P.E., Handley P.S. Bacterial coaggregation: an integral process in the development of multi-species biofilms. TRENDS in Microbiology. 2003. 11(2).pp. 94-100.
- [29] Kolenbrander, P.E. et al. Communication among oral bacteria. Microbiol. Mol. Biol. Rev. 2002. 66.pp. 486-505.
- [30] Gorlov I.F., Lebedev A.T., Galkov V.Y., Orlyanskiy A.V., Shlykov S.N. Effects of feed additives "Yoddar-Zn" and "Glimalask-Vet" on the productivity of beef cattle. Research journal of pharmaceutical biological and chemical sciences. 2016. 7(5) pp. 2518-2522.
- [31] Gorlov, Ivan Fiodorovich; Titov, Evgeniy Ivanovich; Semenov, Gennadiy Viacheslavovich, Slozhenkina, Marina Ivanovna; Sokolov, Aleksandr Yurievich; Omarov, Ruslan Saferbegovich); Goncharov, Aleksandr Ivanovich; Zlobina, Elena Yurievna; Litvinova, Elena Viktorovna; Karpenko, Ekaterina Vladimirovna. INTERNATIONAL JOURNAL OF FOOD PROPERTIES Volume: 21. Issue: 1. 2018. P. 1031-1042.
- [32] Omarov, Ruslan Saferbegovich; Antipova, Lyudmila Vasilevna; Konieva, Oksana Nikolaevna; Meshcheryakov, Vladimir Anatolyevich; Shlykov, Sergei Nikolaevich. Biotechnological Aspects In The Development Of Functional Food Products. Research journal of pharmaceutical biological and chemical sciences. Volume: 9. Issue: 3. P.: 751-755. Publ: MAY-JUN 2018.
- [33] Gorlov, Ivan Fedorovich; Omarov, Ruslan Saferbegovich; Slozhenkina, Marina Ivanovna; Zlobina, Elena Yuryevna; Mosolova, Natalia Ivanovna; Shlykov, Sergei Nikolaevich. Study Of The Influence Of Beef With An Improved Fatty Acid Composition On The Development Of Atherosclerosis In Animal Experiments. Research journal of pharmaceutical biological and chemical sciences Volume: 9 Issue: 4 P: 1159-1162 Publ: JUL-AUG 2018.
- [34] Shlykov, Sergei Nikolayevich, Omarov, Ruslan Saferbegovich. Analyzing Methods For Improving Beef Tenderness. research journal of pharmaceutical biological and chemical sciences Volume: 9. Issue: 4.
 P.: 1135-1137. Publ: JUL-AUG 2018.
- [35] Omarov, Ruslan Saferbegovich; Nesterenko, Anton Alekseyevich; Chimonina, Irina Victorovna; Sangadzhieva, Lyudmila Khalgaevna; Sangadzhieva, Olga Stanislavovna; Shlykov, Sergei Nikolayevich.



Development Of Food Products Enriched With Biologically Active Form Of Iron. Research journal of pharmaceutical biological and chemical Volume: 9. Issue: 4. P.: 902-905. Publ: JUL-AUG 2018.