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A New Antidiabetic Compound 8,9-dimethoxy Ellagic Acid from Sasaladaan (*Peperomia pellucida* L. Kunth)

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ABSTRACT

Peperomia pellucida (L.) Kunth.) is a annual herb belongs to Piperaceae family and also know as sasaladaan or suruhan in Indonesia. These herbs has been known empirically used as blood sugar reducer. Previous research has proved that ethanol extract and ethyl acetat fraction have a good potential as antidiabetics. This study was conducted to obtain antidiabetic compounds of *P. pellucida* using chromatographic techniques guided by in vivo activity test. A new antidiabetic compound, 8,9-dimethoxy ellagic acid (1), was isolated from ethyl acetat fraction of *P. pellucida*. The chemical structure of 1 was determined by spectroscopic methods and comparison with those related compound previously reported. The 8,9-dimethoxy ellagic acid at dose of 100 mg/kg body weight produced significant antidiabetic activity in alloxan-induced hyperglycemic Swiss Webster mice.

Keywords: Antidiabetic, 8,9-dimethoxy-ellagic acid, Peperomia pellucida, Piperaceae.





INTRODUCTION

Diabetes mellitus is the largest endocrine disease worldwide characterized by elevated blood glucose levels and disturbance in carbohydrate, fat and protein metabolism [1]. Diabetic patients experience various vascular complications such as, atherosclerosis, diabetic nephropathy, retinopathy and neuropathy [2]. The currently available therapy for diabetic includes insulin and various oral antidiabetic agents such as the sulfonylureas, biguanides, thiazolidinediones and α -glucosidase inhibitors. Each of the oral antidiabetic agents is however, associated with a number of serious adverse effects [3,4]. Plant-based drugs have been known to be safe and cheaper. Many natural product and herbal medicines have been studied in the search for an effective management of diabetes mellitus and most of them have therapeutic claims [5,6,7]. As part of our continuing search for antidiabetic agents from Indonesian medicinal plant, the ethanolic extract of *Peperomia pellucida* showed a significant antidiabetic activity in alloxan-induced hyperglycemic Wistar rats.

P. pellucida known as "sasaladaan" in Indonesia is a annual herb that typically grows in wet rock crevices, is found from nothest to the south of Indonesia [8]. The plant is used in Indonesian folk medicine for the treatment of fever, headache, contused wound and skin diseases [9]. Previous phytochemical studies on the genus *Peperomia* have revealed the presence of a variety of compounds with interesting biological activities, including flavonoids [10,11], benzopyran derivatives [12,13], secolignans, lignans [14,15,16], A dimeric ArC2 compound, arylpropanoids, phenolic compounds [17,18,19] and essential oils [20]. Although secondary metabolites of other *Peperomia* species have been investigated previously, the antidiabetic compounds from the leaves of *P. pellucida* which growth in Indonesia is yet to be reported. The isolation and structure elucidation of the new antidiabetic compound from the leaves of *P. pellucida* are described herein.

MATERIALS AND METHODS

Plant material: The leaves of *Peperomia pellucida* were collected from Kawangluwuk village, South Congeang, Sumedang, Indonesia in September 2012 and identified by biologist Mr. Joko Kusmoro (Padjadjaran University). A voucher specimen (No. 01/HB-IX/2012) has been deposited at the Herbarium of the Department of Biology, Padjadjaran University, Sumedang, Indonesia.

Experimental animals: This experiment used animal models (mice) *Mus musculus* for antidiabetic activity test that has been approved by the ethics (ethical clearance) of the Health Research Ethics Committee, Faculty of Medicine, University of Padjadjaran, Bandung, West Java, Indonesia. Healthy albino mice of Swiss Webster strain weighing 25-30 g (weight of mice somewhat lower than standard, but the mice were otherwise healthy, 6-8 weeks old, took normal average daily diet, displayed normal daily activity and behavior). Mice of male sex were used, they were obtained from the Central Animal House, Department of Biology, Padjadjaran University. The animals were housed in standar polyprophylene cages and maintained under controlled room temperature ($25 \pm 5^{\circ}$ C) and humidity ($55 \pm 5^{\circ}$) with 12:12 hour light and dark cycle. The mice were fed commercially available mice normal pellet diet and water *ad libitum* [21].

Chemical materials:

Chromatographic separations were carried out on silica gel 60 (Merck). PTLC glass plates were precoated with silica gel GF₂₅₄ (Merck, 0.25 mm). TLC plates were precoated with silica gel GF₂₅₄ (Merck, 0.25 mm). Antidiabetic activity test was performed by *blood glucose level measurement using gluco-test strips* (*FreeStyle Optium H*).

Instruments:

The UV-Visible spectrum was obtained on UV-Shimadzu 1800 series spektrofotometer. Optical rotations were recorded on an ATAGO AP-300 polarimeter. The IR spectra were recorded on a Perkin-Elmer spectrum-100 FT-IR in KBr. Mass spectra were obtained with a Water UP-LC MS/MS instruments. ¹H- and ¹³C-NMR spectra were obtained with a JEOL JNM A-500 spectrometer using TMS as internal standard



Methods:

Procedure 1:

Extraction and isolation: Dried powder leaves (4.4 kg) of *P. pellucida* were extracted by soxhlet apparatus with *n*-hexane, EtOAc, *n*-BuOH and EtOH, successively. Evaporation of these extract under reduced pressure resulted in the crude extracts of *n*-hexane (117 g), EtOAc (125 g), *n*-BuOH (86 g) and EtOH (104 g), respectively. The ethyl acetate extract produced significant antidiabetic activity in alloxan-induced hyperglycemic Wistar rat at dose of 250 mg/kg body weight. A portion of the EtOAc (120 g) was subjected to vacuum liquid chromatography (VLC) using gradient elution of *n*-hexane-EtOAc-MeOH to afford 12 fractions (Ea1-Ea12). Fraction Ea3 (20 g) was subjected to vacuum liquid chromatography (VLC) using gradient elutions (Ea3-1-Ea3-10). Fraction Ea3-1 (3.3 g) was subjected to column chromatography of silica gel, eluted with CH₂Cl₂-EtOAc (9:1) to afford 8 fractions (Ea3-1.1-Ea3-1.8). Fraction Ea3-1.3 (500 mg) was preparative TLC on silica gel GF₂₅₄, eluted with *n*-hexane:EtoAc (7:3) and CH₂Cl₂:EtOAc (9.5:0.5) to give **1** (11 mg).

Procedure 2:

Induction of diabetes: Leaving aside six mice for normal control group, diabetes was induced in 24 mice by a single intraperitoneal injection of alloxan monohydrate in the dose of 245 mg/kg body weight. The fasting blood glucose was determined after 24 hours [21,22]. Eighteen mice showing a blood glucose level of >170 mg/100 mL were taken for the study.

Experimental Design for Antidiabetic Study: A total of 24 animals were equally divided into four groups with six animals in each group [23]:

Group-A: Normal control (Normal saline; 10 mL/kg/d) Group-B: Diabetic control (Normal saline; 10 mL/kg/d) Group-C: Diabetic Test (1; 100 mg/kg/d) Group D: Diabetic Standar (Glibenclamide; 0.7mg/kg/d)

The above drugs were administrated orally once daily for three days. The blood glucose level were measured everyday until the fifth day for each group.

Method of blood glucose estimation: Blood glucose estimation was carried out by Glucose Oxidase Method (GOD-PAP) using blood glucose test strips (FreeStyle Optium H). In order to assess the effect of alloxan and to chemically establish the diabetic condition, an incision was done in any of the four veins in the tail of the mice using scalpel blade 7 days after induction. A sample of the mice's venous blood was collected on a reagent strip 7 days after the diabetes induction procedure for blood glucose level determination using portable glucose analyzer. In this study, mice with glucose levels above 170 mg/dL were considered as having severe diabetes [21].

RESULTS AND DISCUSSION

The dried and powdered leaves of *P. pellucida* was extracted with *n*-hexane, ethyl acetate, *n*-butanol and ethanol succesively. All the extracts were evaluated for antidiabetic activity at dose of 250 mg/kg body weight in alloxan-induced hyperglycemic Wiss albino mice; the ethyl acetate showed strongest antidiabetic activity. By using the antidiabetic assay *in vivo* in alloxan-induced hyperglycemic Swiss Webster albino mice, the ethyl acetate extract was subjected to multiple chromatographic steps, using silica gel G_{60} and preparative TLC to afford a new benzopyran compound (1) (Figure 1).

Compound **1** was obtained as a pale yellowish powder, completely dissolved in methanol and showed flourencence under UV light at λ 384, 329, 278 and 248 nm, $[\alpha]_{20}^{p} + 24.7^{\circ}$ (*c* 0.2, MeOH). The molecular formula, C₁₅H₁₂O₄, was determined from LC-MS/MS (*m/z* 257.3145 [M+H]⁺, calcd for C₁₅H₁₂O₄ 256.3674) and NMR spectrosopic data (Table 1), requiring ten degrees of unsaturation. The UV spectrum showed absorption maximum at λ_{max} 384, 329, and 207 nm, indicated the presence of conjugated double bond. The IR spectrum showed the presence of carbonyl lactone (1718 cm⁻¹), conjugated double bond (1612 cm⁻¹)and ether (1089 cm⁻¹)



¹) functionalities. The ¹H-NMR spectrum (Table 1) showed the presence of two aromatik proton singlet at [δ_{H} 7.15 (1H, s, H-10) and 7.82 (1H, s, H-7)] due to tetrasubstituted benzene ring, four other aromatic protons at [δ_{H} 7.57 (1H, d, *J*=7.2 Hz, H-4), 7.91 (1H, d, *J*=6.7 Hz, H-1), 7.70 (1H, dd, *J*=6.7, 7.8 Hz, H-3) and 9.26 (1H, dd, *J*=7.2, 7.8 Hz, H-2)] from disubstituted benxene ring, and two methoxyl signal at [δ_{H} 4.13 (3H, s) and 4.20 (3H, s)]. In the ¹³C NMR spectrum (Table 1), 15 carbon signals appeared, which were assigned by DEPT and HMQC experiments as one carbonyl lactone (δ_{C} 169.2), six sp² methines (δ_{C} 129.8, 128.3, 128.2, 126.3, 110.4, and 105.4), four sp² quartenary carbons (δ_{C} 127.6, 136.2, 122.7 and 121.5), two quartenary oxyaryl carbons (δ_{C} 152.1 and 155.6) and two methoxyl carbons (δ_{C} 60.5 and 57.2). These functionatilities accounted for seven out of the ten indices of hydrogen deficiency.



Figure 1: Chemical structure of compound 1.

Position	¹³ C NMR, δc	δ H NMR, δ_{H} (integral,
	(mult.)	mult. <i>, J</i> Hz)
1	129.8 (d)	7.91 (1H, d, 6.7)
1a	127.6 (s)	-
2	128.3 (d)	9.26 (1H, dd, 7.2, 7.8)
3	128.2 (d)	7.70 (1H, dd, 6.7, 7.8)
4	126.3 (d)	7.57 (1H, d, 7.2)
4a	136.2 (s)	-
6	169.2 (s)	-
6a	122.7 (s)	-
7	110.4 (d)	7.82 (1H, s)
8	152.1 (s)	-
9	155.6 (s)	-
10	105.4 (d)	7.15 (1H, s)
10a	121.5 (s)	-
11	60.4 (q)	4.13 (3H, s)
12	57.2 (q)	4.20 (3H, s)].

Table 1: NMR data (500 MHz for ¹H and 125 MHz for ¹³C, in asetone-*d*₆)

The remaining three hydrogen deficiency were consistent with benzopyran structure [17,18]. Comparison of the NMR data for **1** with those urolithin A [17-19] indicated that compound **1** was an analogue of this later compound. The main difference is additional methoxyl groups at C-8 and C-9, indicating that compound **1** is 8,9-dimethoxy of ellagic acid. The gross structure of **1** was deduced from ¹H-¹H COSY and HMBC spectra (Figure 2).



Figure 2: Selected ¹H-¹H COSY and HMBC correlations for **1**.

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In the HMBC spectrum, the correlations arising from methoxyl protons at dH 4.13 and 4.20 to C-9 (δ_{C} 155.6), C-10 (δ_{C} 105.4), C-8 (δ_{C} 152.1), and C-7 (δ_{C} 110.4), enable the assignment of the two methoxyl groups was located at C-8 and C-9, respectively. Proton aromatic signals at δ_{H} 7.91 and 9.26 were coupled to each other and were correlated to C-1a (δ_{C} 127.6) and C-10a (δ_{C} 121.5), whereas other proton aromatic signals at δ_{H} 7.57 and 7.70 were coupled to each other and were correlated to C-3 (δ_{C} 128.2) and C-4a (δ_{C} 136.2), suggested that benzopyran ring was located at C-4a, C-1a, C-10a and C-6a. The location of benzopyran ring was supported also from correlation from aromatic signals at δ_{H} 7.82 to C-6a (δ_{C} 122.7) and C-6 (δ_{C} 169.2) and δ_{H} 7.15 to C-10a (δ_{C} 121.5).

Compound 1 was evaluated for its antidiabetic activity in alloxan-induced hyperglycemic mice. Compound 1 exhibited 33.74% (p<0.01) blood glucose lowering in normoglycemic model at dose of 100 mg/kg.

8,9-dimethoxy of ellagic acid (1)

Yellowish amorphous powder. $[\alpha]^{p}_{20} - 24.7^{\circ}$ (*c*, 0.2, MeOH) UV (MeOH) λ max: 384, 329, 278 and 248 nm. IR (KBr) ν_{max} : 1718, 1612, 1089 cm⁻¹ ¹H NMR (acetone-*d*₆, 500 MHz): Table 1 ¹³C NMR (acetone-*d*₆, 125 MHz): Table 1 LC-MS/MS: (positive ion mode), *m/z* 257.3145 [M+H]⁺, calcd. for C₁₅H₁₂O₄, 256.3674).

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CONCLUSIONS

Based above data and biogenetic point of view the occurance benzopyran compound in this genus, consequently compound **1** was determined as a new benzoypran derivative and was named 6H-dibenzo[b,d]-pyrano-6-one-8,9-dimethoxy or 8,9-dimethoxy of ellagic acid. Compound **1** exhibited 33.74% (p<0.01) blood glucose lowering in glycemic model at dose of 100 mg/kg.

REFERENCES

- [1] Aguwa C.N. (2004) Theurapeutic Basis for Clinical Pharmacy in the Tropics, 3rd edition. SNAAP Press Ltd, Enugu. pp. 1-230.
- [2] Sheetz M.J. (2002) Molecular understanding of hyperglycemias adverse effects for diabetic complication. *J. Am. Med. Assoc.* 288, 2579-2588
- [3] Moller D.E. (2001) New Drug Targets for Type 2 Diabetes and Metabolic Syndrome. *Nature* 414,821-825.
- [4] Nwaegerue E., Nweke I.N., Ezeala, C.C, Unekwe P.C. (2007) Glucose lowering effect of leaf extracts of *Viscum album* in normal and diabetic rats. *J. Res. Med. Sci.* 12(5), 235-240.
- [5] Jung M., Park M. Lee H.C., Kang Y.H., Kang E.S., and Kim S. (2006) Antidiabetic Agent from Medicinal Plants. *Curr. Med. Chem.* 13(10):1203-18
- [6] Bnouham, M., Ziyyat, A., Mekfi, H., Tahri, A., and Legssyer, A. (2006) Medicinal plants with potential activity-A review of ten years of herbal medicine reserch (1990-2000). *Int. J. Diabetes & Metabolism*, 141-25
- [7] Rao, M.U., Sreenivasulu, M., Chengaiah, B., Reddy, K.J., and Chetty, C.M. (2010) Herbal medicines for diabetes mellitus : A Review, *Int. J. of Pharm.Tech. Research*, 2(3): 1882-1892.
- [8] Heyne, K. (1987) *Tumbuhan Berguna Indonesia II.* Badan Penelitian dan Pengembangan Kehutanan. Yayasan Sarana Wana Jaya. Jakarta. 462
- [9] Kasahara, S. (1995) *Medicinal Herb Index in Indonesia*. 2nd ed. PT. Eisai Indonesia, 21
- [10] Mota, J.S., Leite, A.C. Kato, M.J., Young, M.C.M., Bolzani, V.S., and Furlan, M. (2011) Isoswertisin flavones and other constituens from *Peperomia obtusifolia*, *Nat. Prod. Res.* 25(1):1-7.



- [11] Velozo, L.S.M., Ferreira, M.C.P., Santos, M.I.S., Moreira, D.L., Guimares, E.F., Emerenciano, V.P., and Kaplan, M.A.C. (2009) C-glycosyl flavones from *Peperomia blanda*, *Fitoterapia* 80, 119-122.
- [12] Seeram, N.P. Jacobs, H., McLean, S., and Reynolds, W.F. (1998) A prenilated benzopyran derivative from *Peperomia clusiifolia*. *Phytochemistry* 49(5):1389-1391.
- [13] Salazar, K.J.M., Guillermo, E., Paredes, D., Lluncor, L.R., Young, M.C.M., and Kato, M.J (2005) Chromenes of polyketide origin from *Peperomia villipetiola*, *Phytochemistry* 66, 573-579.
- [14] Wu, J.L, Li, N., Hasegawa, T. Sakai, Mitsui, T., Oguru, H., Kataoka, T., Oka, S., Kiuchi, M, Tomida, A., Tsuruo, T., Li, M., Tang, W., and Ando, M. (2006) Bioactive secolignans from *Peperomia dindygulensis*, *J. Nat. Prod.* 69, 790-794
- [15] Zhang, G.L., Li, Na, Wang, Y.H., Zheng, Y.T., Zhang, Z., and Wang, M.W. (2007) Bioactive lignans from *Peperomia heyneana*, *J. Nat. Prod.* 70, 662-664.
- [16] Zu, Y.H., Li, Na, and Wang, M.W. (2008) A new lignan glycoside from *Peperomia duclouxii*, *Nat. Prod. Res.*, 22 (17): 1483-1486.
- [17] Bayma, J.C., Arruda, M.S.P., Muller, A.H. Arruda, A.C., and Canto, W.C. (2000) A dimeric ArC2 compound from *Peperomia pellucida*, *Phytochemistry* 55, 779-782.
- [18] Tanaka, T., Asai, F., and linuma, M (1998) Phenolic compounds from *Peperomia obtusifolia*, *Phytochemistry* 49(1): 229-232.
- [19] Xu, S., Li N., Ning, M.M, Zhou, C.H., Yang, Q.R, and Wang, M.W (2006) Bioactive compounds from *Peperomia pellucida*, *J. Nat. Prod.* 69, 247-250
- [20] Da Silva, M.H., Zoghbi, M.G.B., Andrade, E.H.A., and Maia, J.G.S. (1999) The essential oils of *Peperomia pellucida* Kunth. and *P. circinnata* Link var. circinnata, *Flav. and Fragr. J.* 14, 312-314.
- [21] Carvalho E.N, Carvalho NAS, Ferreira LM (2003) Expeerimental model of induction of diabetes mellitus in rats. *Acta Cir Bras* [Serial online] 18. Special edition. Available on <u>URL:http://www.scielo.br/acb</u>.
- [22] Das, S. and Barman, S., 2012. Antidiabetic and antihyperlipidemic effects of ethanolic extract of leaves of *Punica granatum* in alloxan-induced non-insulin-dependent diabetes mellitus albino rats. *Indian J. Pharmacol.* 44(2): 219-224
- [23] Sheikh, H., Sikder, S., Paul, S.K., A.M. Hasan, R., Rahanam, M.M., and Kundu, S.P. 2012. Hypoglicemic, anti-inflammatory and analgesic activity of *Peperomia pellucida*(L.) HBK (Piperaceae)., *IJPSR* 4(1):458-463.