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Chemical Constituents of Anacolosa frutescens.

Agnes B Alimboyoguen¹, Kathlia A De Castro-Cruz¹, Chien-Chang Shen², and Consolacion Y Ragasa^{3,4*}.

¹School of Chemical Engineering and Chemistry, Mapúa Institute of Technology, Muralla St., Intramuros, Manila 1002 Philippines.

²National Research Institute of Chinese Medicine, 155-1, Li-Nong St., Sec. 2, Taipei 112, Taiwan.

³Chemistry Department, De La Salle University Science & Technology Complex Leandro V. Locsin Campus, Biñan City, Laguna 4024, Philippines.

⁴Chemistry Department, De La Salle University, 2401 Taft Avenue, Manila, 1004 Philippines.

ABSTRACT

Chemical investigation of the dichloromethane extract of the leaves of *Anacolosa frutescens* afforded 3-acetylaleuritolic acid (1), β -amyrin (2) and a mixture of monounsaturated and saturated fatty acids. The structures of 1 and 2 were identified by comparison of their ¹³C NMR data with those reported in the literature. **Keywords**: *Anacolosa frutescens*, Olacaceae, 3-acetylaleuritolic acid, β -amyrin

*Corresponding author

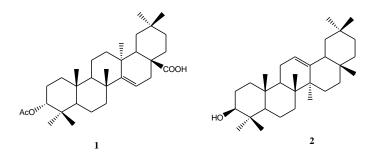
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INTRODUCTION

Anacolosa frutescens (Blume) Blume, commonly known as galonut, is a shrub or small tree which is a native of tropical Asia [3]. The fruits including the seeds are used as food [1], while the wood is used for house posts [4]. There are no reported studies on the chemical constituents and biological activities of *A. frutescens*. However, its congener, *A. pervilleana* was recently studied and found to exhibit antiviral properties. *A. pervilleana* afforded anacolosine, octadeca-9,11,13-triynoic acid, (13*E*)-octadec-13-en-9,11-diynoic acid, (13*E*)-octadec-13-en-11-ynoic acid, lupenone, β -amyrone, and (*S*)-sambunigrin. Lupenone and β -amyrone showed a moderate activity against CHIKV (EC₅₀ 77 and 86 μ M, respectively) and the acetylenic acids exhibited IC₅₀ values around 3 μ M in the DENV RdRp assay [3].

We report herein the isolation and identification of the triterpenes, 3-acetylaleuritolic acid (1), β amyrin (2), and a mixture of monounsaturated and saturated fatty acids from the leaves of *A. frutescens*. To the best of our knowledge this is the first report on the isolation of these compounds from *A. frutescens*.



MATERIALS AND METHODS

General experimental procedures

NMR spectra were recorded on a Varian VNMRS spectrometer in CDCl₃ at 600 MHz for ¹H NMR and 150 MHz for ¹³C NMR spectra. Column chromatography was performed with silica gel 60 (70-230 mesh). Thin layer chromatography was performed with plastic backed plates coated with silica gel F_{254} and the plates were visualized by spraying with vanillin/H₂SO₄ solution followed by warming.

Sample Collection

The leaves of the plant were collected at Baranggay Tambo Balagbag, Indang, Cavite in January, 2013. The leaves were identified as *Anacolosa frutescens* (Blume) Blume at the Jose Vera Santos Herbarium, Institute of Biology, University of the Philippines, Diliman, Quezon City.

Isolation

The air-dried leaves (1.22 kg) of *A. frutescens* were ground in a blender, soaked in CH_2Cl_2 for three days and then filtered. The filtrate was concentrated under vacuum to afford a crude extract (65.9 g) which was chromatograped using increasing proportions of acetone in CH_2Cl_2 at 10% increment. The 20% acetone in CH_2Cl_2 fraction was rechromatographed (8 ×) using petroleum ether to afford a mixture of monounsaturated and saturated fatty acids (8 mg). The 40% acetone in CH_2Cl_2 fraction was rechromatographed (6 ×) using 10% EtOAc in petroleum ether to yield **2** (5 mg) after washing with 2.5% EtOAc in petroleum ether. The 50% acetone in CH_2Cl_2 fraction was rechromatographed (5 ×) using 15% EtOAc in petroleum ether, followed by 20% EtOAc in petroleum ether (2 ×) to afford **1** (3 mg).

3-Acetylaleuritolic acid (1)

Colorless solid. ¹³C NMR (150 MHz, CDCl₃): δ 37.37 (C-1), 23.45 (C-2), 80.00 (C-3), 37.66 (C-4), 55.58 (C-5), 18.69 (C-6), 40.77 (C-7), 39.00 (C-8), 49.04 (C-9), 37.91 (C-10), 17.30 (C-11), 33.65 (C-12), 37.31 (C-13), 160.59 (C-14), 116.72 (C-15), 31.36 (C-16), 51.33 (C-17), 41.48 (C-18), 35.32 (C-19), 29.27 (C-20), 33.31 (C-21), 30.71 (C-22),



27.94 (C-23), 16.58 (C-24), 15.61 (C-25), 26.13 (C-26), 22.39 (C-27), 183.56 (C-28), 31.89 (C-29), 28.68 (C-30), 171.00, 21.30 (OAc).

β-Amyrin (2)

Colorless solid. ¹³C NMR: 38.77 (C-1), 27.22 (C-2), 79.03 (C-3), 38.57 (C-4), 55.15 (C-5), 18.37 (C-6), 32.49 (C-7), 39.78 (C-8), 47.62 (C-9), 36.94 (C-10), 23.52 (C-11), 121.71 (C-12), 145.19 (C-13), 41.71 (C-14), 26.14 (C-15), 26.14 (C-16), 32.64 (C-17), 47.22 (C-18), 46.82 (C-19), 31.08 (C-20), 34.72 (C-21), 37.13 (C-22), 28.09 (C-23), 15.49 (C-24), 15.58 (C-25), 16.79 (C-26), 25.99 (C-27), 28.39 (C-28), 33.33 (C-29), 23.68 (C-30).

RESULTS AND DISCUSSION

Silica gel chromatography of the dichloromethane extract of the leaves of *A. frutescens* afforded **1-2** and a mixture of monounsaturated and saturated fatty acids. Their structures were identified by comparison of their ¹H and ¹³C NMR data with those reported in the literature for 3-acetylaleuritolic acid (1) [4], β -amyrin (2) [5] and a mixture of monounsaturated and saturated fatty acids.

Triterpene **1** exhibited antimicrobial activity against *S. aureus* and *S. typhimurium* [6]; significant inhibitory activity on vitality of adult male worms of *O. gutturosa* [7]; strong inhibition of DNA topoisomerase II and high cytotoxicity against human lung carcinoma A549 cells [8]. On the other hand, β -amyrin (**2**) was reported to possess anti-inflammatory activity [9, 10] and analgesic property [11, 12].

REFERENCES

- [1] Verheij EWM, Coronel RE, eds. Edible fruits and nuts. 2:64 In: Faridah Hanum, I. & L. J. G. van der Maesen, eds., Plant Resources of South-East Asia (PROSEA). 1991; 2: 64.
- [2] Tipot L. Forest Research Institute Malaysia. 1995; 272: 274–275.
- [3] Bourjot M, Leyssen P, Eydoux C, Guillemot J-C, Canard B, Rasoanaivo P, Guéritte F, Litaudon N. Fitoterapia 2012; 83(6): 1076–1080.
- [4] Alimboyoguen AB, De Castro-Cruz KA, Shen C-C, Li W-T, Ragasa CY. J Chem Pharm Res 2014; 6(5): 1318-1320.
- [5] Raga DD, Alimboyoguen AB, Shen C-C, Ragasa CY. Philipp Agric Scient 2011; 94(2): 103-110.
- [6] Peres MY, Delle Monache F, Cruz AB, Pizzolatti MG, Yunes RAJ. J Ethnopharmacol 1997; 56: 223-226.
- [7] Nyasse B, Ngantchou I, Nono JJ, Schneider B. Nat Prod Res 2006; 20: 391-397.
- [8] Wada S, Tanaka R. Chem Biodivers 2006; 3: 473-479.
- [9] Recio MC, Giner RM, Manez S, Rios JL. Planta Med 1995; 61(2): 181-185.
- [10] Madeiros R, Otuki MF, Avellar MC, Calixto JB. Eur J Pharmacol 2007; 559: 227-235.
- [11] Otuki C, Ferreira J, Lima F, Meyre-Silva C, Malheiros A, Muller L, Cani G, Santos A, Yunes R, Calixto J. J Pharmacol Exp Therap 2005; 31(1): 310-318.
- [12] Soldi C, Pizzolatti G, Luiz A, Marcon R, Meotti F, Miotob L, Santos A. Bioorganic and Medicinal Chemistry 2008; 16(6): 3377-3386.