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Triterpenes and Sterols from *Samanea saman*.

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ABSTRACT

The dichloromethane extract of *Samanea saman* afforded epilupeol (**1**), lupenone (**3**) and chlorophyll a (**3**) from the leaves; **2** and lupeol (**4**) from the peduncle; and **4**, unsaturated triglycerides (**5**), α -spinasterol (**6**), and α -spinasterone (**7**) from the twigs. The structures of **1-6** were identified by comparison of their ¹H and/or ¹³C NMR data with those reported in the literature.

Keywords: *Samanea saman*, Fabaceae, epilupeol, lupenone, lupeol, α -spinasterol, α -spinasterone

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INTRODUCTION

Samanea saman (Jacq.) Merr. of the family Fabaceae is commonly known as acacia or rain tree. In the Philippines, it is widely planted as a shade tree. A decoction of the bark and leaves is used to treat diarrhea, acute bacillary dysentery, enteritis, colds, sore throat and headache. A decoction of fresh material is applied as external wash for anaphylactic dermatitis, eczema, skin pruritus [1]. *S. saman* was reported to exhibit potent antimicrobial, molluscicidal, nematocidal, hemolytic, and hypercholesterolemic properties [2]. Literature search on the chemical constituents of *S. saman* revealed the presence of octacosanol, α -spinasterol, β -D-glucose of α -spinasterol, kaempferol and pithecolobine from the different parts of the tree [3]. The volatile constituents of *S. saman* have been reported with palmitic acid (55.5%), 1,8-cineole (15.9%), and oleic acid (7.4%) as the major constituents [2, 4]. Another study reported the isolation of lupeol and epilupeol from the whole plant of *S. saman* [5].

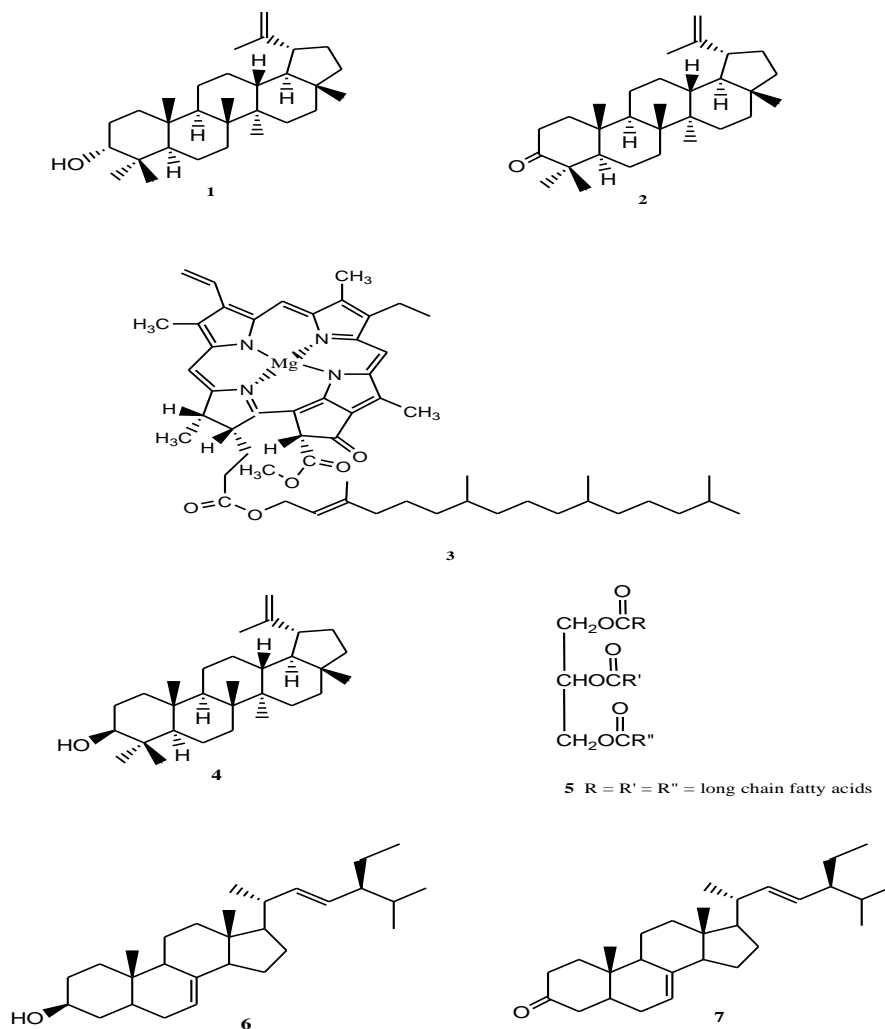


Fig. 1. Chemical constituents of *Samanea saman*: epilupeol (1), lupenone (2), chlorophyll a (3), lupeol (4), unsaturated triglycerides (5), α -spinasterol (6), and α -spinasterone (7).

This study was conducted as part of our research on the chemical constituents of trees found at the De La Salle University–Manila campus. We earlier reported the chemical constituents of *Barringtonia asiatica* [6-7], *Alstonia scholaris* [8], *Pterocarpus indicus* [9-10], and *Swietenia macrophylla* [11]. In this study, the isolation and identification of epilupeol (**1**), lupenone (**2**) and chlorophyll a (**3**) from the leaves; **2** and lupeol (**4**) from the peduncle; and **4**, unsaturated triglycerides (**5**), spinasterol (**6**), and spinasterone (**7**) from the twigs of *S. saman* are reported.

MATERIALS AND METHODS

General Experimental Procedures

NMR spectra were recorded on a Varian VNMRS spectrometer in CDCl_3 at 600 MHz for ^1H NMR and 150 MHz for ^{13}C NMR spectra. Column chromatography was performed with silica gel 60 (70-230 mesh), while the TLC was performed with plastic-backed plates coated with silica gel F₂₅₄. The plates were visualized with vanillin- H_2SO_4 and warming.

A glass column (18 inches in height and 1.0 inch internal diameter) was packed with silica gel. The crude extract was fractionated by silica gel chromatography using increasing proportions of acetone in dichloromethane (10 % increments) as eluents. 100 mL fractions were collected. All fractions were monitored by thin layer chromatography. Fractions with spots of the same *Rf* values were combined and rechromatographed. A glass column (12 inches in height and 0.5 inch internal diameter) was used for the rechromatography. 5mL fractions were collected. Final purifications were conducted using Pasteur pipettes as columns. 1 mL fractions were collected.

Sample Collection

The sample was collected from the De La Salle University-Manila Campus in July 2013. It was identified as *Samanea saman* (Jacq.) Merr. at the Bureau of Plant Industry, Manila, Philippines.

Isolation of Chemical Constituents

The air-dried leaves (264 g), petioles (43 g) and twigs (165 g) of *S. saman* were separately ground in a blender, soaked in CH_2Cl_2 for three days and then filtered to afford crude extracts: leaves (4.34 g), petioles (0.42 g) and stems (1.56 g). The crude extracts were separately fractionated by silica gel chromatography using increasing proportions of acetone in dichloromethane (10 % increments) as eluents.

The 10% acetone in CH_2Cl_2 fractions from the chromatography of the crude leaves extract was rechromatographed (4 ×) in 5% EtOAc in petroleum ether to afford **2** (4 mg) after washing with petroleum ether. The 20% to 30% acetone in CH_2Cl_2 fractions from the chromatography of the crude petioles extract were combined and rechromatographed (3 ×) in

10% EtOAc in petroleum ether afford **1** (6 mg) after washing with petroleum ether. The 40% to 50% acetone in CH₂Cl₂ fractions from the chromatography of the crude leaves extract were combined and rechromatographed (4 ×) in 15% EtOAc in petroleum ether to afford **3** (4 mg) after washing with petroleum ether, followed by Et₂O.

The 30% to 50% acetone in CH₂Cl₂ fractions from the chromatography of the crude petioles extract were combined and rechromatographed (3 ×) in 10% EtOAc in petroleum ether, followed by 12.5% EtOAc in petroleum ether. The fractions eluted with 10% EtOAc in petroleum ether were combined and rechromatographed (2 ×) in the same solvent to afford **4** (2 mg) after washing with petroleum ether. The fractions eluted with 12.5% EtOAc in petroleum ether were combined and rechromatographed (3 ×) in the same solvent to afford **2** (3 mg) after washing with petroleum ether.

The 10% acetone in CH₂Cl₂ fractions from the chromatography of the crude twigs extract was rechromatographed (3 ×) in 5% EtOAc in petroleum ether to afford **7** (2 mg) after washing with petroleum ether. The 20% acetone in CH₂Cl₂ fractions from the chromatography of the crude twigs extract was rechromatographed (5 ×) in 5% EtOAc in petroleum ether to afford **5** (3 mg). The 30% to 40% acetone in CH₂Cl₂ fractions from the chromatography of the crude twigs extract were combined and rechromatographed (2 ×) in 10% EtOAc in petroleum ether, followed by 12.5% EtOAc in petroleum ether. The fractions eluted with 10% EtOAc in petroleum ether were combined and rechromatographed (3 ×) in the same solvent to afford **4** (4 mg) after washing with petroleum ether. The fractions eluted with 12.5% EtOAc in petroleum ether were combined and rechromatographed (4 ×) in the same solvent to afford **6** (3 mg) after washing with petroleum ether.

Epilupeol (1): ¹³C NMR (150 MHz, CDCl₃): δ 33.24 (C-1), 25.39 (C-2), 76.26 (C-3), 37.52 (C-4), 49.02 (C-5), 18.27 (C-6), 34.13 (C-7), 41.02 (C-8), 50.20 (C-9), 37.28 (C-10), 20.77 (C-11), 25.11 (C-12), 38.01 (C-13), 42.90 (C-14), 27.37 (C-15), 35.58 (C-16), 43.01 (C-17), 48.23 (C-18), 48.03 (C-19), 151.04 (C-20), 29.84 (C-21), 40.00 (C-22), 28.24 (C-23), 22.13 (C-24), 15.91 (C-25), 15.96 (C-26), 14.62 (C-27), 18.00 (C-28), 109.29 (C-29), 19.28 (C-30).

Lupenone (2): ¹³C NMR (150 MHz, CDCl₃): δ 39.61 (C-1), 34.16 (C-2), 218.23 (C-3), 47.33 (C-4), 54.91 (C-5), 19.67 (C-6), 33.55 (C-7), 40.77 (C-8), 49.78 (C-9), 36.87 (C-10), 21.46 (C-11), 25.14 (C-12), 38.16 (C-13), 42.89 (C-14), 27.42 (C-15), 35.51 (C-16), 47.98 (C-17), 48.23 (C-18), 47.95 (C-19), 150.89 (C-20), 29.82 (C-21), 39.97 (C-22), 26.64 (C-23), 21.03 (C-24), 15.77 (C-25), 15.97 (C-26), 14.47 (C-27), 18.00 (C-28), 109.38 (C-29), 19.30 (C-30).

Lupeol (4): ¹³C NMR (150 MHz, CDCl₃): δ 38.70 (C-1), 27.42 (C-2), 79.01 (C-3), 38.86 (C-4), 55.29 (C-5), 18.32 (C-6), 34.28 (C-7), 40.83 (C-8), 50.43 (C-9), 37.17 (C-10), 20.92 (C-11), 25.14 (C-12), 38.05 (C-13), 42.83 (C-14), 27.44 (C-15), 35.58 (C-16), 43.00 (C-17), 47.99 (C-18), 48.30 (C-19), 150.99 (C-20), 29.85 (C-21), 40.00 (C-22), 27.98 (C-23), 15.36 (C-24), 16.11 (C-25), 15.97 (C-26), 14.54 (C-27), 18.00 (C-28), 109.31 (C-29), 19.30 (C-30).

Spinasterol (6): ^{13}C NMR (150 MHz, CDCl_3): δ 37.13 (C-1), 31.46 (C-2), 71.06 (C-3), 37.98 (C-4), 40.24 (C-5), 29.65 (C-6), 117.45 (C-7), 139.56 (C-8), 49.43 (C-9), 34.21 (C-10), 21.54 (C-11), 39.54 (C-12), 43.28 (C-13), 55.11 (C-14), 23.01 (C-15), 28.51 (C-16), 55.88 (C-17), 12.04 (C-18), 13.04 (C-19), 40.83 (C-20), 21.37 (C-21), 138.17 (C-22), 129.42 (C-23), 51.24 (C-24), 31.92 (C-25), 21.09 (C-26), 19.02 (C-27), 25.40 (C-28), 12.25 (C-29).

RESULTS AND DISCUSSION

The dichloromethane extract of *Samanea saman* afforded epilupeol (**1**) [12], lupenone (**2**) [13] and chlorophyll a (**3**) [14] from the leaves; **2** and lupeol (**4**) [15] from the peduncles; and **4**, unsaturated triglycerides (**5**) [16], α -spinasterol (**6**) [17], and α -spinasterone (**7**) [18] from the twigs. The structures of **1-7** were identified by comparison of their ^1H and/or ^{13}C NMR data with those reported in the literature [12-18].

Although bioassays were not conducted on the isolated compounds, there were previous studies that reported on their biological activities.

A mixture of lupenone (**2**) and caryophyllene oxide in a 1:4 ratio showed *in-vitro* typanocidal activity against epimastigotes forms of *T. cruzi* ($\text{IC}_{50} = 10.4 \mu\text{g}/\text{mL}$, $\text{FIC} = 0.46$) [19]. Triterpene **2** from *E. multiflora* stimulated melanogenesis in B16 murine melanoma cells through the inhibition of ERK1/2 activation, indicating that it can be used as a possible treatment for hypopigmentation diseases [20].

Lupeol (**4**) exhibited anticancer activities against pancreatic [21], prostate [22-23], ovarian [24], colorectal and myeloma [25], breast [26], stomach [27], cervical [26-28], lymphoma [28], leukemia [26, 29], melanoma and neuroblastoma [29], melanoma [25-27, 29-30], and lung [25-28, 30] cancers. Furthermore, **4** was also found to exhibit antimicrobial [31], anti-inflammatory [32], and anti-arthritis [33-34] properties.

α -Spinasterol (**6**) exhibited antiproliferative action against CACO-2 cell line with IC_{50} value of 60 nM/ml [35]. Moreover, **6** has significant therapeutic potential to modulate the development and/or progression of diabetic nephropathy [36]. It was also found to exhibit anti-angiogenic potential [37]. It was also reported to exhibit antioxidative [38], antinociceptive [39], anti-inflammatory [40], anti-ulcerogenic [41] and antitumor [42] effects.

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REFERENCES

- [1] Philippines Medicinal Plants. Medicinal Uses for Samanea Saman/National Tropical Botanical Garden, May, 2014. available at <http://www.stuartxchange.com/Acacia.html.com>
- [2] Isika OA, Tameka WM, William SN, Essien E. Afr J Biotech 2006; 5(20): 1890-1893.
- [3] Misra G, Nigam SK, Mira CR. Phytochem 1971; 10: 3313-3314.
- [4] Isika OA, Tameka WM, William SN. Nat Prod Commun 2007; 2(12): 1314.
- [5] Ferdous F, Hossain MdK, Rahman MS, Hossain MdA, Kabir S, Rashid MA. Dhaka Univ J Pharm Sci 2010; 9(2): 69-73.
- [6] Ragasa CY, Espineli DL, Shen C-C. Chem Pharm Bull 2011; 59(6): 778-782.
- [7] Ragasa CY, Espineli DL, Shen C-C. Nat Prod Res 2012; 26(20): 1869-1875.
- [8] Ragasa CY, Lim F, Raga DD, Shen C-C. Pharm Chem J 2013; 47(1): 54-58.
- [9] Ragasa CY, Hofileña JG, de Luna RD. Nat Prod Res 2005; 19(4): 305-309.
- [10] Hofileña JG, Ragasa CY. Kimika 2002; 18(1): 5-8.
- [11] Ragasa CY, Daniel R, Rideout JA. Kimika 1998; 14(1): 21-25.
- [12] Mohd Lip J, Nazrul Hisham D, Arif Zaidi J, Musa Y, Ahmad AW, Normah A, Sharizan A. J Trop Agric Food Sci 2009; 37(2): 195-201.
- [13] Tsai P-W, de Castro-Cruz K, Shen C-C, Ragasa CY. Phcog J 2012; 4(31): 1-4.
- [14] Ragasa CY, de Jesus J. Res. J Pharm Biol Chem Sci 2014; 5(3):
- [15] Ragasa CY, Lim F, Raga DD, Shen C-C. Pharm Chem J 2013; 47(1): 54-58.
- [16] Ragasa CY, Lorena GS, Mandia EH, Raga DD, Shen C-C. Amer J Essent Oils Nat Prod 2013; 1(2): 7-10.
- [17] Raga DD, Herrera AA, Alimboyoguen AB, Shen C-C, Ragasa CY. Philipp Agric Scient 2011; 94(2):103-110.
- [18] Kihisa TA, Kimura Y, Tai T, Arai K. Chem Pharm Bull 1999; 47(8): 1161-1163.
- [19] Polanco-Hernández G, Escalante-Erosa F, García-Sosa K, Rosado ME, Guzmán-Marín E, Acosta-Viana KY, Giménez-Turba A, Salamanca E, Peña-Rodríguez LM. Evid-Based Complement Alter Med 2013; Article ID 435398, 6 pages, <http://dx.doi.org/10.1155/2013/435398>.
- [20] Villareal MO, Han J, Matsuyama K, Sekii Y, Smaoui A, Shigemori H, Isoda H. Planta Med 2013; 79(3-4): 236-43.
- [21] Murtaza I, Saleem N, Adhami VM, Hafeez BB, Mukhtar H. Cancer Res 2009; 69: 1156-1165.
- [22] Saleem M, Murtaza I, Tarapore RS, Suh Y, Adhami VM, Johnson JJ, Siddiqui IA, Khan N, Asim M, Hafeez BB, Shekhani MT, Li B, Mukhtar H. Carcinog 2009; 30: 808-817.
- [23] Prasad S, Nigam S, Kalra N, Shikla Y. Mol Carcinog 2008; 47: 916-924.
- [24] Chaturvedula VSP, Schilling JK, Miller JS, Andriantsiferona R, Rasamison VE, Kingston DGI. J Nat Prod 2004; 67: 895-898.
- [25] Gauthier C, Legault J, Lebrum M, Dufour P, Pichetel A. Bioorg Med Chem 2006; 14: 6713-6725.
- [26] Cmoch P, Pakulski Z, Swaczynova J, Strnad M. Carbohydr Res 2008; 343: 995-1003.
- [27] Hata K, Hori K, Murata J, Takahashi S. J Biochem 2005; 138: 467-472.
- [28] Lin L, Chou C, Kuo Y. J Nat Prod 2001; 64: 674-676.

- [29] Hata K, Hori K, Ogasawara H, Takahashi S. *Toxicol Lett* 2003; 143: 1-7.
- [30] You YJ, Nam NH, Kim Y, Bae KH, Ahn BZ. *Phytother Res* 2003; 17: 341-344.
- [31] Shai LJ, McGraw LJ, Adea MA, Mdee LK, Eloff JN. *J Ethnopharmacol* 2008; 119: 238-244.
- [32] Recio MC, Giner RH, Manez S, Rios JC. *Planta Med* 1995; 61: 182-185.
- [33] Latha RM, Lenin M, Rasool M, Varalakimi P. *Prostag Leukotr Ess* 2001; 64: 81-85.
- [34] Kweifio-Okai G, Field B, Rumble BA, Macrides TA, de Munk F. *Drug Dev Res* 1995; 35: 137-141.
- [35] Ravikumar YS, Mahadevan KM, Manjunatha H, Satyanarayana ND. *Phytomed* 2009; 17(7): 513-518.
- [36] Jeong SI, Kim KJ, Choi MK, Keum KS, Lee S, Ahn SH, Back SH, Song JH, Ju YS, Choi BK, Jung KY. *Planta Med* 2004; 70(8): 736-9.
- [37] Raga DD, Alimboyoguen AB, Shen C-C, Herrera AA, Ragasa CY. *Philipp Agric Scient* 2011; 94: 103-110.
- [38] Coballase-Urrutia E, Pedraza-Chaverri J, Camacho-Carranza R, Cárdenas-Rodríguez N, Huerta-Gertrudis B, Medina-Campos ON, Mendoza-Cruz M, Delgado-Lamas G, Espinosa-Aguirre JJ. *Toxicol* 2010; 276: 41-48.
- [39] Meotti FC, Ardenghi JV, Pretto JB, Souza MM, d' Avila Moura J, Junior AC, Soldi C, Pizzolatti MG, Santos AR. *J Pharm Pharmacol* 2006; 58: 107-112.
- [40] Boller S, Soldi C, Marques MC, Santos EP, Cabrini DA, Pizzolati MG, Zampronio AR, Otuki MF. *J Ethnopharmacol* 2010; 130: 262-266.
- [41] Klein Jr LC, Gandolfi RB, Santin JR, Lemos M, Cechinel Filho V, de Andrade SF. *Naunyn Schmiedebergs Arch Pharmacol* 2010; 381: 121-126.
- [42] Jeon GC, Park MS, Yoon DY, Shin CH, Sin HS, Um SJ. *Exp Molec Med* 2005; 37: 111-120.