# Research Journal of Pharmaceutical, Biological and Chemical Sciences 

# Isolation and Characterization of Polygonum equisetiforme Flavonoids and Their Acaricidal Activity against Tetranychus urticae Koch. 

Abdel-Aziz M Dawidar ${ }^{1 *}$, Mamdouh Abdel-Mogib ${ }^{1}$, Mahmoud E El-Naggar ${ }^{2}$, and Mohamed E Mostafa².<br>${ }^{1}$ Chemistry Dept., Fac. of Sci., Mansoura Univ., Mansoura, Egypt.<br>${ }^{2}$ Plant Protection Research Institute, Agriculture Research Center, Egypt.

## ABSTRACT

Acaricidal activity assessment against Tetranychus urticae Koch (Acari: Tetranychidae) of different extracts of Polygonum equisetiforme (Polygonaceae) aerial parts revealed that butanol extract has the highest activity. Processing of this fraction using repeated column and thin layer chromatographic techniques resulted in the isolation of five flavonoids, which have identified NMR as; quercetin (1), avicularin (2), isoquercitrin(3), betmidin (4) and myricitrin (5). All the five isolated flavonoids showed acaricidal activity using leaf-dip technique against $T$. urticae Koch larvae in the following decreasing activity order; quercetin ( $\mathrm{LC}_{50}=11.28 \mathrm{ppm}$ ), avicularin ( 46.72 ppm ), isoquercitrin ( 63.77 ppm ), myricitrin ( 169.66 ppm ) and betmidin ( 188.11 ppm ). While the activity against the adult females was found to be quercetin ( $\mathrm{LC}_{50}=16.28 \mathrm{ppm}$ ), isoquercitrin ( 93.38 ppm ), avicularin ( 98.57 ppm ), betmidin ( 161.00 ppm ) and myricitrin ( 206.68 ppm ) after 7 days of treatment. This is the first report on isolation of the two flavonoids betmidin (4) and myricitrin (5) from $P$. equisetiforme.
Keywords: Acaricidal activity, Tetranychus urticae Koch, polygonum equisetiforme, polygonaceae, Flavonoids, quercetin, isoquercitrin, avicularin, betmidin, myricitrin.

## *Corresponding author

## INTRODUCTION

Tetranychus urticae Koch is a worldwide pest, feeding on a large variety of plant families in Egypt, attacking cotton, fruit trees and vegetables. This mite showed readily and rapidly resistance to synthetic acaricides, moreover, synthetic acaricides comprises high toxic impact towards mammals and birds and unfriendly to environment. Therefore, it is recommended to search for natural acaricides from plants, which has stimulated us to examine some available Egyptian plants for these desired botanicals [1-3].

Polygonum equisetiforme (Polygonaceae) was used as a cure lotion for treating poisonous snakebites in ancient Egypt. [4] In addition, it has anti-inflammatory, astringent therapeutically effects [5] and its roots water extract was applied for treating kidney and urinary tract diseases [6].

Ethanol extracts from $P$. equisetiforme exhibited strong antimicrobial activity against Gram-positive bacteria [7, 8]. Polysaccharides were isolated from roots of P. equisetiforme found to be effective natural antioxidants, with little antitumor activity. [9] Previous phytochemical investigation of Polygonum equisetiforme (Polygonaceae) revealed the isolation of quercetin, quercetin-3-O-rhamnoside, quercetin-3-O-glucuronide, quercetin-3-O-arabinoside and isorhamnetin [7]. In addition the isolation and identification of apigenin-6-C-arabino-pyranosyl-8-C-glucopyranoside, apigenin-6,8 di C-gluco-pyranoside, quercetin-3-O-glucopyranoside, quercetin-3,7-di-O-gluco-pyranoside, rhamnetin-3-O-rhamno-pyranoside, myricetin ( $3,5,7,3^{\prime}, 4^{\prime}, 5^{\prime}$-hexahyroxyflavone) and gallic acid were reported [10]. We report here the flavonoids isolated from Polygonum equisetiforme as natural acaricides against T. urticae Koch.

## MATERIALS AND METHODS

## General

NMR experiments were performed on a Bruker AMX 400 instrument standard pulse sequences operating at 400 MHz for ${ }^{1} \mathrm{H}-\mathrm{NMR}$ and 100 MHz for ${ }^{13} \mathrm{C}-\mathrm{NMR}$. Chemical shifts are given in $\delta(\mathrm{ppm})$ relative to TMS as internal standard material and the coupling constants (J) are in Hz. UV spectra were recorded on Shimadzu, model 2401.

## Chemicals

Thin layer chromatography and preparative TLC were performed on silica gel (Kieselgel 60, F 254) of 0.25 mm thickness. Pet. ether $\left(60-80^{\circ} \mathrm{C}\right)$, methylene chloride, ethyl acetate and methanol were obtained from Adwic Company.

## Plant material

The aerial parts of Polygonum equisetiforme was collected from Al-Aresh city, Sinai at April, 2009, identified by Prof. Dr. Loutfy Boulos, Professor of Botany, Faculty of Science, Alexandria University, Egypt.

## Extraction and isolation

The dried powdered aerial parts of the plant ( 1.15 kg ) were extracted by a soxhlet apparatus using pet. ether, methylene chloride, ethyl acetate and methanol, successively. The solvents were evaporated under vacuum to give pet. ether fraction ( 7.79 g ), methylene chloride fraction ( 5.78 g ), ethyl acetate fraction ( 6.32 g ) and methanol fraction ( 116.75 g ), which was further washed by butanol to give the butanol fraction ( 4.35 g ). Butanol fraction ( 4.35 g ) was subjected to Sephadex LH-20 CC using MeOH as solvent to give ten subfractions. The third subfraction has been applied on TLC Silica gel plates using EtOAc-MeOH- $\mathrm{H}_{2} \mathrm{O}$ (98:1:1) as a developing system to afford compound (3), ( $26 \mathrm{mg}, \mathrm{R}_{\mathrm{f}} 0.13$ ). The fourth subfraction was purified on silica gel TLC, developed by the same previous system to afford compound (2), ( $42 \mathrm{mg}, \mathrm{R}_{\mathrm{f}}$ 0.54 ), (4), ( $12 \mathrm{mg}, R_{f} 0.33$ ) and (5), ( $14 \mathrm{mg}, R_{f} 0.20$ ). The fifth subfraction has been separated on TLC Silica gel plates using EtOAc to give compound (1) ( $59 \mathrm{mg}, \mathrm{R}_{\mathrm{f}} 0.85$ ).

Quercetin 3-O- $\alpha$-L-arabinofuranoside (Avicularin) (2). Yellow needle crystals. $\mathrm{R}_{\mathrm{f}} 0.54$; UV ( $\mathrm{MeOH}, \max$ ) 256, $358 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}, \delta, \mathrm{ppm}, \mathrm{J}, \mathrm{Hz}$ ): 7.54 (br s, H-2'), 7.51 (br d, J = 8.3, H-6'), 6.92 (br d, J = 8.3, H-5'), 6.41 (br s, H-8), 6.23 (br s, H-6), 5.48 (br s, H-1'), 4.35 (br d, J = 1.3, H-2'), 3.95 (dd, J = 3.8, 8.3, H-4'), 3.93 (m, H-3'), $3.55\left(\mathrm{~m}, 2 \mathrm{H}-5^{\prime \prime}\right) ;{ }^{13} \mathrm{C}$ NMR ( 100 MHz, ס, ppm): 179.9 (C-4), 165.9 (C-7), 163.0 (C-5), 159.3 (C-2), 158.5 (C-9), 149.8 (C-4'), 146.3 (C-3'), 134.9 (C-3), 123.0 (C-1'), 122.9 (C-6'), 116.4 (C-2'), 116.8 (C-5'), 109.5 (C-1'), 105.6 (C-10), 99.8 (C-6), 94.7 (C-8), 88.0 (C-4"), 83.2 (C-2'), 78.6 (C-3"), 62.5 (C-5").

Quercetin 3-O - $\beta$-D-glucopyranoside (Isoquercitrin) (3). Yellow powder. $\mathrm{R}_{\mathrm{f}} 0.13$; UV (MeOH) 258, $360 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}, \delta, \mathrm{ppm}, \mathrm{J}, \mathrm{Hz}$ ): 7.86 (d, J = 1.7, H-2'), 7.59 (dd, J = 1.7, 8.4, H-6'), 6.87 (d, J = 8.4, H-5'), 6.36 (br s, H-8), 6.18 (br s, H-6), 5.12 (d, J = 7.8, H-1'), 3.65 (dd, J = 5.1,11.1, H-6"), 3.57 (dd, J = 5.1,11.1, H-6"), 3.83 (m, H-2"), 3.58 (m, H 3'), 3.87 (m, H-4"), 3.51 ( $\mathrm{m}, \mathrm{H}-5^{\prime \prime}$ ); ${ }^{13} \mathrm{C}$ NMR ( $100 \mathrm{MHz}, \delta, \mathrm{ppm}$ ): 179.2 (C-4), 168.3 (C-7), 162.8 (C-5), 158.6 (C-2), $158.5(\mathrm{C}-9), 149.9$ (C-4'), 145.8 (C-3'), 135.7 (C-3), 122.9 (C-1'), 122.8 (C-6'), 117.7 (C-5'), 116.1 (C-2'), 105.6 (C-10), 104.9 (C-1"), 100.6 (C-6), 95.2 (C-8), 77.1 (C-5"), 75.1 (C-3"), 73.2 (C-2'), 70.0 (C-4"), 61.9 (C-6").

Myricetin 3-O- $\alpha$-L-arabinofuranoside (Betmidin) (4). Yellow powder. $\mathrm{R}_{\mathrm{f}} 0.33$; UV (MeOH) 256, 354 nm ; ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}, \delta, \mathrm{ppm}$, J, Hz): 7.15 (br s, H-2'), 7.15 (br s, H-6'), 6.40 (d, J = 2, H-8), 6.22 (d, J = 2, H-6), 5.48 (br s, H-1'), 4.21 (br d, J = 1.8, H-2"), 3.95 (m, H-3"), 3.95 (m, H$\left.4^{\prime \prime}\right), 3.51\left(\mathrm{t}, \mathrm{J}=3.4,2 \mathrm{H}-5^{\prime \prime}\right)$; ${ }^{13} \mathrm{C}$ NMR ( $100 \mathrm{MHz}, \delta, \mathrm{ppm}$ ): 179.9 (C-4), 166.0 (C-7), 163.0 (C-5), 159.3 (C-2), 158.5 (C-9), 146.8 (C-3'), 146.8(C-5'), 137.9 (C-4'), 134.8 (C-3), 121.9 (C-1'), 109.4 (C$\left.6^{\prime}\right), 109.4$ (C-2'), 109.4 (C-1'), 105.4 (C-10), 99.9 (C-6), 94.8 (C-8), 88.0 (C-4"), 83.2 (C-2"), 78.7 (C-3"), 62.5 (C-5").

Myricetin 3-O- $\alpha$-L-rhamnopyranoside (Myricitrin) (5). Yellow powder. $\mathrm{R}_{\mathrm{f}} 0.20$; UV (MeOH) 256, $350 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CD}_{3} \mathrm{OD}, \delta, \mathrm{ppm}, \mathrm{J}, \mathrm{Hz}$ ): 6.97 (br s, H-2'), 6.97 (br s, H-6'), 6.37 (d, J = 2, H-8), 6.21 (d, J = 2, H-6), 5.33 (d, J = 1.1, H-1'), 4.21 ( $\mathrm{m}, \mathrm{H}-2^{\prime \prime}$ ), 3.81 ( $\left.\mathrm{dd}^{\prime}, \mathrm{J}=3.1,9.4, \mathrm{H}-3^{\prime \prime}\right)$, 3.51 (dq, J = 6.1, 9.4, H-5"), 3.33 ( t , J = 9.4, H-4"), 0.98 (d, J = 6.2, H-6"); ${ }^{13} \mathrm{C}$ NMR ( $100 \mathrm{MHz}, \delta$, ppm): 179.6 (C-4), 166. 0 (C-7), 163.1 (C-5), 159.3 (C-2), 158.5 (C-9), 146.8 (C-3'), 146.8(C-5'),
137.8 (C-4'), 136.2 (C-3), 121.9 (C-1'), 109.5 (C-6'), 109.5 (C-2'), 105.7 (C-10), 103.6 (C-1'), 99.8 (C-6), 94.7 (C-8), 73.3 (C-4''), 72.0 (C-3'), 72.0 (C-5''), 71.8 (C-2'), 17.6 (C-6').

## Maintance of spider mite colony

Colony of spider mite Tetranychus urticae Koch was reared under laboratory condition $\left(25 \pm 2{ }^{\circ} \mathrm{C}\right.$ and $60 \pm 5 \%$ R.H) at Plant Protection Research Institute branch, Dakahlia Governorate. This study colony was isolated from heavily infested castor oil plant leaves. Spider mite colony was reared on castor oil leaves. These leaves were cleaned and placed on moisten cotton wool pad in Petri dishes. This colony was left for one year under the precious conditions in order to get a homogenous and sensitive colony. Spider mite individuals were transferred to the leaves by aid of fine camels hair brush. Breeding leaves were changed twice weekly at summer and once weekly at winter. Adding water was done twice daily to prevent escaping of $T$. urticae individuals.

## Assessment of acaricidal activity

In this respect, laboratory experiments are conducted to evaluate the activity of various tested plant extracts against $T$. urticae mobile stages (larvae and adult females). The leaf-dip technique was used [11].

The indication of mortality was chosen as the failure of mites to respond positively by leg movement followed light brooding with a fine brush. Mortality percentages were determined and corrected by using Abotts (1925) formula and they are statistically analyzed to estimate $\mathrm{LC}_{50}, \mathrm{LC}_{90}$ and slope values according to Finney (1971) [12, 13]. Toxicity index was computed for different extracts and the isolated compounds by comparing these materials with the most effective extracts or isolated compounds using Sun's (1950) equation [14].

$$
\text { Toxicity index }=\frac{\mathrm{LC}_{50} \text { of compound } \mathrm{A}}{\mathrm{LC}_{50} \text { of compound } \mathrm{B}}
$$

Where:
A is the most effective compound
$B$ is the tested compound

## RESULTS AND DISCUSSION

Processing of the plant aerial parts has resulted in four fractions; pet. ether, methylene chloride, ethyl acetate and butanol fractions (cf. Experimental). The acaricidal activity indicated that butanol fraction was the most effective fraction against the larvae and adult females of $T$. urticae after 7-days of treatment that guided and stimulated us to search for its bioactive components. Repeated column and thin layer chromatography has resulted in the isolation of five flavonoids. They were characterized by NMR spectroscopy (cf. Experimental).

Examination of NMR spectra of the five isolated compounds from the butanol fraction in addition to their UV absorption spectra indicated that they belong to flavonoids. The ${ }^{1} \mathrm{H}-\mathrm{NMR}$ data of compound (1) indicated that ring A is 5,7-disubstituted, as shown by two meta-located protons at $\delta 6.15 \mathrm{ppm}(1 \mathrm{H}, \mathrm{br}, \mathrm{H}-6)$ and $\delta 6.39 \mathrm{ppm}(1 \mathrm{H}, \mathrm{br} \mathrm{s}, \mathrm{H}-8)$. On the other hand, the observation of $A B X$ system at $\delta 7.63 \mathrm{ppm}\left(1 \mathrm{H}, \mathrm{brs}, \mathrm{H}-2^{\prime}\right), \delta 6.85 \mathrm{ppm}\left(1 \mathrm{H}, \mathrm{br} \mathrm{d}, \mathrm{J}=8.4 \mathrm{~Hz}, \mathrm{H}-5^{\prime}\right)$ and $\delta 7.50 \mathrm{ppm}\left(1 \mathrm{H}, \mathrm{dd}, \mathrm{J}=2.3,8.4 \mathrm{~Hz}, \mathrm{H}-6^{\prime}\right)$ has suggested a $3^{\prime},^{\prime}$-disubstituted ring B , which was identified as quercetin [11].


Quercetin (1)


Isoquercitrin (3)


Avicularin (2)


Betmidin (4)


Myricitrin (5)

A 3-O-substituted quercetin structures were indicated for compounds (2) and (3) due to the corresponding anomeric protons at $\delta 5.48 \mathrm{ppm}(1 \mathrm{H}, \mathrm{br} \mathrm{s})$ and $\delta 5.12 \mathrm{ppm}(1 \mathrm{H}, \mathrm{d}, \mathrm{J}=7.8 \mathrm{~Hz})$, characteristic for arabinofuranosyl and glucopyranosyl moieties, respectively (Table 1). The structure of compounds (2) and (3) were characterized as quercetin 3-O- $\alpha$-L-arabinofuranoside (avicularin) and quercetin 3-O- $\beta$-D-glucopyranoside (isoquercitrin) by H-H COSY, HSQC and HMBC and comparison with literature data [15].

Table 1: NMR data ( $\delta \mathrm{ppm}$ ) for compounds 2-5.

| Position | (2) |  | (3) |  | (4) |  | (5) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\begin{gathered} 1 \mathrm{H} \\ \text { (multiplicity, J) } \end{gathered}$ | ${ }^{13} \mathrm{C}$ | $\begin{gathered} 1 \mathrm{H} \\ \text { (multiplicity, J) } \end{gathered}$ | ${ }^{13} \mathrm{C}$ | $\begin{gathered} 1 \mathrm{H} \\ \text { (multiplicity, J) } \end{gathered}$ | ${ }^{13} \mathrm{C}$ | $\begin{gathered} 1 \mathrm{H} \\ \text { (multiplicity, J) } \end{gathered}$ | ${ }^{13} \mathrm{C}$ |
| 2 | - | 159.31 | - | 158.59 | - | 159.35 | - | 159.39 |
| 3 | - | 134.9 | - | 135.71 | - | 134.86 | - | 136.27 |
| 4 | - | 179.97 | - | 179.22 | - | 179.99 | - | 179.62 |
| 5 | - | 163.03 | - | 162.84 | - | 163.03 | - | 163.17 |
| 6 | 6.23 (1H, brs) | 99.87 | 6.18 (1H, brs) | 100.65 | $\begin{gathered} 6.22(1 \mathrm{H}, \mathrm{~d}, \\ \mathrm{J}=2 \mathrm{~Hz}) \end{gathered}$ | 99.99 | $\begin{gathered} 6.21(1 \mathrm{H}, \mathrm{~d}, \mathrm{~J}=2 \\ \mathrm{Hz}) \end{gathered}$ | 99.87 |
| 7 | - | 165.99 | - | 168.33 | - | 166.40 | - | 166.08 |
| 8 | 6.41(1H, brs) | 94.76 | 6.36(1H, brs) | 95.25 | $\begin{gathered} \hline 6.40(1 \mathrm{H}, \mathrm{~d}, \\ \mathrm{J}=2 \mathrm{~Hz}) \end{gathered}$ | 94.82 | $\begin{gathered} 6.37(1 \mathrm{H}, \mathrm{~d}, \mathrm{~J}=2 \\ \mathrm{Hz}) \end{gathered}$ | 94.73 |
| 9 | - | 158.53 | - | 158.49 | - | 158.58 | - | 158.50 |
| 10 | - | 105.61 | - | 105.64 | - | 105.46 | - | 105.79 |
| $1{ }^{\prime}$ | - | 123.08 | - | 122.90 | - | 121.99 | - | 121.91 |
| $2^{\prime}$ | 7.54 (1H, brs) | 116.43 | $\begin{gathered} 7.86(1 \mathrm{H}, \mathrm{~d}, \\ \mathrm{J}=1.7 \mathrm{~Hz}) \\ \hline \end{gathered}$ | 116.07 | 7.15 (1H, s) | 109.41 | 6.97 (1H, s) | 109.57 |
| 3' | - | 146.32 | - | 145.82 | - | 146.85 | - | 146.83 |
| 4' | - | 149.82 | - | 149.97 | - | 137.98 | - | 137.88 |
| $5^{\prime}$ | $\begin{gathered} 6.92(1 \mathrm{H}, \mathrm{brd}, \\ \mathrm{J}=8.3 \mathrm{~Hz}) \end{gathered}$ | 116.83 | $\begin{gathered} 6.87(1 \mathrm{H}, \mathrm{brd}, \\ \mathrm{J}=8.4 \mathrm{~Hz}) \end{gathered}$ | 117.71 | - | 146.85 | - | 146.83 |
| $6^{\prime}$ | $\begin{gathered} 7.51(1 \mathrm{H}, \mathrm{brd}, \\ \mathrm{J}=8.3 \mathrm{~Hz}) \\ \hline \end{gathered}$ | 122.96 | $\begin{aligned} & 7.59(1 \mathrm{H}, \mathrm{dd}, \\ & \mathrm{J}=1.7,8.4 \mathrm{~Hz}) \end{aligned}$ | 122.88 | 7.15 (1H, s) | 109.41 | 6.97 (1H, s) | 109.57 |
| $1{ }^{\prime \prime}$ | 5.48 (1H, brs) | 109.53 | $\begin{gathered} 5.12(1 \mathrm{H}, \mathrm{~d}, \\ \mathrm{J}=7.8 \mathrm{~Hz}) \end{gathered}$ | 104.96 | 5.48 (1H, brs) | 109.41 | $\begin{gathered} 5.33(1 \mathrm{H}, \mathrm{~d}, \\ \mathrm{J}=1.1 \mathrm{~Hz}, \end{gathered}$ | 103.60 |
| 2" | $\begin{gathered} 4.35(1 \mathrm{H}, \text { brd, } \\ \mathrm{J}=1.3 \mathrm{~Hz}) \\ \hline \end{gathered}$ | 83.27 | 3.83 (1H, m) | 73.18 | $\begin{gathered} 4.21(1 \mathrm{H}, \text { brd, } \\ \mathrm{J}=1.8 \mathrm{~Hz}) \\ \hline \end{gathered}$ | 83.24 | 4.21 (1H, m) | 71.87 |
| $3{ }^{\prime \prime}$ | 3.93 (1H, m) | 78.68 | 3.58 (1H, m) | 75.13 | 3.95 (1H, m) | 78.76 | $\begin{aligned} & 3.81(1 \mathrm{H}, \mathrm{dd}, \\ & \mathrm{J}=9.4,3.1 \mathrm{~Hz}) \end{aligned}$ | 72.11 |
| 4" | $\begin{aligned} & 3.95(1 \mathrm{H}, \mathrm{dd}, \\ & \mathrm{J}=8.3,3.8 \mathrm{~Hz}) \end{aligned}$ | 88.02 | 3.87 (1H, m) | 70.01 | 3.95 (1H, m) | 88.05 | $\begin{gathered} 3.33(1 \mathrm{H}, \mathrm{t}, \\ \mathrm{J}=9.5 \mathrm{~Hz}) \end{gathered}$ | 73.34 |
| 5" | 3.55 (2H, m) | 62.53 | 3.51 (1H, m) | 77.14 | $\begin{gathered} 3.51(2 \mathrm{H}, \mathrm{t}, \\ \mathrm{J}=3.4 \mathrm{~Hz}) \end{gathered}$ | 62.56 | $\begin{aligned} & 3.51(1 \mathrm{H}, \mathrm{dq}, \\ & \mathrm{J}=9.4,6.1 \mathrm{~Hz}) \end{aligned}$ | 72.02 |
| 6"a | - | - | $\begin{gathered} 3.65(1 \mathrm{H}, \mathrm{dd}, \\ \mathrm{J}=5.1,11.1 \mathrm{~Hz}) \end{gathered}$ | 61.92 | - | - | $\begin{gathered} 0.98(3 \mathrm{H}, \mathrm{~d}, \\ \mathrm{J}=6.2 \mathrm{~Hz}) \end{gathered}$ | 17.66 |
| 6"b |  |  | $\begin{gathered} 3.57(1 \mathrm{H}, \mathrm{dd}, \\ \mathrm{J}=5.1,11.1 \mathrm{~Hz}) \end{gathered}$ |  |  |  |  |  |

Compounds (4) and (5) were found to belong to myricetin glycosides, as established by the typical UV absorption ( $\lambda_{\max } 256,350 \mathrm{~nm}$ ) and NMR spectra (Table 1). The sugar residues were determined by the characteristic sugar anomeric proton signals at $\delta 5.48 \mathrm{ppm}(1 \mathrm{H}, \mathrm{br}$ s) and $\delta 5.33 \mathrm{ppm}(1 \mathrm{H}, \mathrm{d}, \mathrm{J}=1.1 \mathrm{~Hz})$, which were assigned to arabinofuranosyl, and rhamnosyl moieties, respectively. Therefore, compounds (4) and (5) were identified as myricetin 3-O-
$\alpha$-L-arabinofuranoside (betmidin) and myricetin 3-O- $\alpha$-L-rhamnopyranoside (myricitrin), respectively. They were confirmed by H-H COSY, HSQC and HMBC and comparison with literature data [16].

It is worthwhile mentioning here, that this is the first report of betmidin (4) and myricitrin (5) from $P$. equisetiforme (Polygonaceae).

## Toxicity effect of the plant fractions to larvae and adult females of Tetranychus urticae (Koch) after 7-days of treatment

The susceptibility of the larvae and adult females of $T$. urticae to various solvent plant extract fractions after 7-days of treatments is reported in table (2). The descending order of the acaricidal activity of both larvae and adult females at $\mathrm{LC}_{50}$ and $\mathrm{LC}_{90}$ levels were butanol fraction, ethyl acetate fraction, methylene chloride fraction and pet. ether fraction. The slopes of the toxicity lines were calculated to be fluctuated and increased from 0.919 in the ethyl acetate fraction to 1.625 in the pet. ether fraction in case of larvae and 1.161 in ethyl acetate fraction to 1.858 in the pet. ether fraction for the adult females. The other fractions lines came between these two fraction lines.

The toxicity index of the $\mathrm{LC}_{50}$ values, as shown from table (2), indicated that butanol fraction was the most effective plant fraction against larvae and adult females of $T$. urticae after 7-days of treatment, followed by ethyl acetate fraction, methylene chloride fraction while pet. ether fraction was the least effective one.

## Toxicity effect of the isolated flavonoids to larvae and adult females of Tetranychus urticae (Koch) after 7-days of treatment

Table (3) showed the susceptibility of both larvae and adult females of $T$. urticae to the isolated flavonoids (1-5). Data revealed that quercetin (1) exhibited the highest efficiency against both larvae and adult females after 7-days of treatment and this was proved from the toxicity index of the isolated flavonoids.

It is most likely from the examination of the chemical structure of the isolated flavonoids and their toxicity index at $\mathrm{LC}_{50}$ that substitution of the hydrogen atom at position $3^{\prime}$ in ring $B$ decreases the acaricidal activity.

Table 2: Toxicity of plant fractions against larvae and adult females of $T$. urticae after 7-days of treatment.

| Plant extract | Larvae |  |  |  | Adult females |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathrm{LC}_{50}$ (ppm) and confidence limits at 95\% | $\mathrm{LC}_{90}$ (ppm) and confidence limits at 95\% | Slope | $\begin{aligned} & \text { Toxicity } \\ & \text { index at } L C_{50} \\ & \text { value } \end{aligned}$ | $\mathrm{LC}_{50}$ (ppm) and confidence limits at 95\% | $\mathrm{LC}_{90}$ (ppm) and confidence limits at 95\% | Slope | Toxicity index at $\mathrm{LC}_{50}$ value |
| Pet. ether fraction | $\begin{gathered} \hline 101.60 \\ 20.49 \quad 172.89 \end{gathered}$ | $\begin{gathered} 624.85 \\ 412.21 \quad 1718.29 \end{gathered}$ | $1.625 \pm 0.464$ | 8.63 | $$ | $\begin{gathered} 1467.90 \\ 999.94 \quad 3405.00 \end{gathered}$ | $1.858 \pm 0.449$ | 6.30 |
| Methylene Chloride fraction | $\begin{array}{cc} \hline 98.72 \\ 17.78 \quad 170.10 \end{array}$ | $\begin{gathered} 894.81 \\ 516.635101 .58 \end{gathered}$ | $1.339 \pm 0.403$ | 8.88 | $\begin{gathered} 129.50 \\ 24.76 \quad 218.51 \end{gathered}$ | $\begin{gathered} 1575.56 \\ 763.87 \quad 25017.60 \end{gathered}$ | $1.181 \pm 0.376$ | 14.58 |
| Ethyl acetate fraction | $\begin{gathered} 23.73 \\ 6.37 \quad 44.60 \end{gathered}$ | $\begin{gathered} 588.61 \\ 294.20 \quad 2677.46 \end{gathered}$ | $0.919 \pm 0.204$ | 36.93 | $$ | $\begin{gathered} 527.59 \\ 269.22 \quad 2665.92 \end{gathered}$ | $1.161 \pm 0.270$ | 45.43 |
| Butanol fraction | $$ | $$ | $0.995 \pm 0.173$ | 100.00 | $$ | $$ | $1.349 \pm 0.408$ | 100.00 |

Table 3: Toxicity of isolated compounds against larvae and adult females of $T$. urticae after 7-days of treatment.

| Isolated | Larvae |  |  |  | Adult females |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Compounds | $\mathrm{LC}_{50}$ (ppm) and confidence limits at 95\% | $\mathrm{LC}_{90}$ (ppm) and confidence limits at 95\% | Slope | Toxicity index at $\mathrm{LC}_{50}$ value | $\mathrm{LC}_{50}$ (ppm) and confidence limits at 95\% | $\mathrm{LC}_{90}$ (ppm) and confidence limits at 95\% | Slope | Toxicity index at $\mathrm{LC}_{50}$ value |
| Quercetin <br> (1) | $$ | $$ | $1.397 \pm 0.400$ | 100.00 | $\begin{gathered} 16.28 \\ 7.29 \quad 22.95 \end{gathered}$ | $\begin{gathered} 81.07 \\ 46.83 \quad 663.37 \end{gathered}$ | $1.838 \pm 0.582$ | 100.00 |
| Avicularin <br> (2) | $$ | $$ | $0.977 \pm 0.236$ | 24.15 | $$ | $\begin{gathered} 995.50 \\ 552.62 \quad 7443.61 \end{gathered}$ | $1.276 \pm 0.397$ | 16.51 |
| Isoquercitrin <br> (3) | $\begin{array}{cc} \hline 63.7688 \\ 11.31 \quad 110.10 \end{array}$ | $\begin{array}{cc} \hline 764.60 \\ 372.79 \quad 11456.51 \end{array}$ | $1.188 \pm 0.378$ | 17.69 | $$ | $$ | $1.068 \pm 0.267$ | 17.43 |
| Betmidin <br> (4) | $$ | $\begin{gathered} \hline 1580.04 \\ 832.7010278 .24 \end{gathered}$ | $1.387 \pm 0.374$ | 6.00 | $$ | $\begin{gathered} \hline 1418.64 \\ 775.49 \quad 5026.86 \end{gathered}$ | $1.356 \pm 0.270$ | 10.11 |
| Myricitrin <br> (5) | $\begin{gathered} \hline 169.66 \\ 109.90239 .17 \end{gathered}$ | $$ | $1.454 \pm 0.274$ | 6.65 | $\begin{gathered} \hline 206.68 \\ 126.61 \quad 314.64 \end{gathered}$ | $\begin{gathered} \hline 2432.75 \\ 1124.1714791 .31 \end{gathered}$ | $1.197 \pm 0.261$ | 7.88 |

## REFERENCES

[1] Dawidar AM, Abdel-Mogib M, El-Naggar ME, Mostafa ME. Rev. Latinoamer. Quím 2009; 37: 14-24.
[2] Attia S, Grissa KL, Ghrabi ZG, Mailleux AC, Lognay G, Hance T. The J Essential Oil Res 2012; 24: 279-288.
[3] Bernardi D, Botton M, Silva da Cunha U, Bernardi O, Malausa T, Garcia MS, Nava DE. Pest Manag Sci 2013; 69: 75-80.
[4] Granat Y. "Medicinal plants of the Negev--Medicinal plants in the Hellenistic period," Environmental Education Publishing House: Sde Boker Academy (1994).
[5] Al-Qura'n S. American J Environ Sci 2005; 1: 74-80.
[6] Said O, Khalil K, Fulder S, Azaizeh H. J Ethnopharmacol 2002; 83: 251-265.
[7] Gazal SA, Abuzarqa M, Mahasneh AM. Phytother Res 1992; 6: 265-269.
[8] Bogdadi HAA, Kokoska L, Havlik J, Kloucek P, Rada V, Vorisek K. Pharm Biol 2007; 45: 386-391.
[9] Ibrahim TA, El-Hela AA. Int J Pharm Bio Sci 2012; 3: 478-492.
[10] Hussein S, EL-Magly U, Tantawy M, Kawashty S, Saleh N. Arabian J Chem 2012; http://dx.doi.org/10.1016/j.arabjc.2012.06.002.
[11] Dittrich V. J Econ Entomol 1969; 55: 633-648.
[12] Abbott WS. J Econ Entomol 1925; 18: 625-627.
[13] Finney DJ. Probit Analysis. A Statistical treatment of the sigmoid Response curve. $7^{\text {th }}$ Ed. Cambridge Univ. Press, Cambridge, England, 1971.
[14] Sun YP. J Econ Entomol 1950; 43: 45-53.
[15] Park B, Matsuta T, Kanazawa T, Park C, Chang K, Onjo M. Chem Natl Comp 2012; 48: 477-479.
[16] Shen C, Chen C, Lee S. J Chinese Chem Soc 2009; 56: 1002-1009.

