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Phytoplankton Bio-Indicators of Water Quality Situations in the Iyagbe Lagoon, South-Western Nigeria.

Onyema IC\*

Department of Marine Sciences, University of Lagos, Akoka, Lagos, Nigeria.

#### **ABSTRACT**

An investigation into the use of phytoplankton species as bio-diagonistic tools and in relation to associated water quality conditions were carried out from October, 2004 to September, 2006 for the lyagbe lagoon in Southwestern Nigeria. Water chemistry conditions ranged from fresh, through brackish to sea situations. Other water chemical parameters showed marked variations and trends. For instance salinity ranged from 1.06 – 35.1‰. The phytoplankton spectrum (76 species) was represented by six divisions namely Bacillariophyta (diatoms, 38 taxa), Cyanophyta (blue-green algae, 18 taxa), Chlorophyta (green algae, 10 taxa), Euglenophyta (euglenoid, 4 taxa), Pyrrophyta (dinoflagellate, 3 taxa) and Chrysophyta (chrysophytes, 2 taxa). Diatoms formed the dominant group and represented a wider array of conditions than any other group. Water quality characteristics reflected by the phytoplankton crop in this study include levels for salinity, pH, cations, depth, nutrients and pollution. Further exactitudes on the bio-diagnostic characteristics of specific organisms are detailed within.

Keywords: Water chemistry, creek, lagoon, lyagbe, algae, pollution, mangrove.

\*Corresponding author

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#### INTRODUCTION

Biological indicators are species used to monitor or assess the health and environmental integrity of an ecosystem (Odiete, 1999). For instance, indicator organisms in perturbed ecosystems are known to provide valuable information with regard to water quality peculiarities of aquatic systems. Changes in water quality could be reflected by the type of species present, abundant, absent or their distributive pattern. According to Onyema (2007a) planktonic microalgae satisfy conditions to qualify as suitable indicators in that they are simple, capable of quantifying changes in water quality, applicable over large geographic areas and can also furnish data on background conditions and natural variability.

Lagoons in south-western Nigeria are generally of two types namely oligohaline and mesohaline lagoons. A few reports exist with regard to available literature on the use of phytoplankton as bio-diagnostic components in assessing water quality status in lagoons of South-western, Nigeria. For instance where as Nwankwo and Akinsoji (1989) investigated and provided a list of pollution tolerant species in an eutrophic creek, Nwankwo and Akinsoji (1992) recorded few species reflecting notable water quality situations in some aquatic systems and associated to the water hyacinth in South-western Nigeria. Furthermore, Nwankwo (2004b) gave record of a number of blue-green algae indicative of a range of polluted scenarios and reported 28 species. Nwankwo *et al.*, (2003) also reported 16 species as harmful algal species for the region. More recently, Onyema and Nwankwo (2006) enunciated 134 species from a polluted tidal creek where as Onyema (2007b) and Onyema and Nwankwo (2009) reported at least 49 and 19 species respectively from suspected polluted aquatic systems in the Lagos area.

Apart from the Lagos lagoon which is probably the most studied of the 10 lagoons in the region, little is known of the physico-chemical variability and phytoplankton diversity and distribution in some other coastal waters, lagoons and their adjoining creeks. Presently only four phycological reports exist for the lyagbe lagoon, namely Onyema (2008) on a checklist of species and Onyema and Nwankwo, (2009) on seasonal primary production level, Onyema (2010) on the diversity and distribution of phytoplankton and Onyema (2013a) on the nutrients and primary production parameters within the estuarine system Additionally however, no single material of the aforementioned acts as a compendium of phytoplankton bio-indicator species in the area at present.

The aim of this investigation was to document the indigenous phytoplankton bio-indicator species of a range of water quality situations in the lyagbe lagoon in south-western Nigeria. This is a first step towards phytoplankton bio-mapping for the region.

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# Study site.

The Iyagbe lagoon (Fig 1) is located in Lagos state, Nigeria and is one of the ten lagoons in South-western Nigeria (Onyema, 2013b). The Porto-Novo and Badagry creeks are the main creeks that make up the Iyagbe lagoon system. For this study 12 stations were chosen as sampling points within the lagoon. The Porto-Novo creek arm of the lagoon is deeper when compared to the Badagry creek. However, the lagoon was generally shallow and is open all year round via the Lagos harbour to the sea. Most locations around the lagoon shore area are devoid of any human presence or activities and are largely rural. The Iyagbe lagoon is a micro-tidal environment. Like all parts of South-western Nigeria, the lagoon is exposed to two distinct seasons namely the

**MATERIALS AND METHODS** 

Like all parts of South-western Nigeria, the lagoon is exposed to two distinct seasons namely the wet (May – October) and the dry (November – April) (Nwankwo, 2004b; Onyema, 2009c). Dense rain forest zone vegetation preceded by littoral mangrove assemblages is the common macrofloral assemblages especially in areas with reduced anthropogenic influence(s).

## **Collection of samples.**

## **Collection of water samples**

The twelve sampling stations were selected to cover the lagoon area and for the collection of samples. Table 1 shows the G.P.S. location, names and number of sampling stations. Monthly surface water samples was collected for twenty-four consecutive months (October, 2004 – September, 2006) for physico-chemical characteristics analysis using 500ml plastic containers with screw caps. Collection of samples from the stations was between 10 and 15hr each time. Water samples were collected just a few centimeters below the water surface at each of the twelve stations. The plastic containers was then labeled appropriately and transported to the laboratory immediately after collection for further analysis.

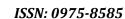
Table 1: G.P.S. location and station name of sampled areas in the lyagbe lagoon.

Station No.	Station name	G.P.S. locations
Station 1	Calabash Island	Latitude 6o 251.987 N, Longitude 3° 231.400 E
Station 2	Tin-can Island	Latitude 6° 251 .833 N, Longitude 3° 211 .532 E
Station 3	Ibafon	Latitude 6° 251 .964 N, Longitude 3° 191 .244 E
Station 4	Imore	Latitude 6° 251 .755 N, Longitude 3° 191 .915 E
Station 5	Ito-ogba	Latitude 6° 251 .409 N, Longitude 3° 141 .624 E
Station 6	Abule-oshun	Latitude 6° 261 .134 N, Longitude 3° 131 .224 E
Station 7	Idiagbon / Igbolobi	Latitude 6° 261.214 N, Longitude 3° 111 .826 E
Station 8	lyagbe	Latitude 6° 251 .603 N, Longitude 3° 111 .990 E

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Station 9	Agbaja	Latitude 6° 241 .473 N, Longitude 3° 121 .744 E
Station 10	Ikare	Latitude 6° 241 .632 N, Longitude 3° 131 .705 E
Station 11	llashe	Latitude 6° 241 .676 N, Longitude 3° 161 .938 E
Station 12	Idimangoro	Latitude 6° 241 .717 N, Longitude 3° 191 .307 E

# Collection of phytoplankton samples.

Phytoplankton sample was collected on each occasion and station with a  $55\mu m$  mesh size standard plankton net towed from a motorized boat for 5mins at low speed (<4 knots). The net was hauled in and the sample transferred into a 250 ml well labeled plastic container with screw cap. Each sample was preserved with 4% unbuffered formalin and stored in the laboratory. After 48hours and prior to microscope analysis, samples were concentrated to 10 ml (Onyema *et al.*, 2007).

## **Biological analysis**

In the laboratory one drop of the concentrated sample, five different times for each sample was investigated at different magnifications (X100 and X400) using a Wild M11 binocular microscope with a calibrated eye piece. The microtransect drop count method, adapted by Onyema (2007a) was used. Appropriate texts were used to aid identification (Smith 1950; Hendey, 1958, 1964; Desikachary, 1959; Wimpenny, 1966; Patrick and Reimer, 1966, 1975; Whitford and Schmacher, 1973; Vanlandingham, 1982; Nwankwo, 1984, 1990, 2004a; Bettrons and Castrejon, 1999; Siver, 2003; Rosowski, 2003).



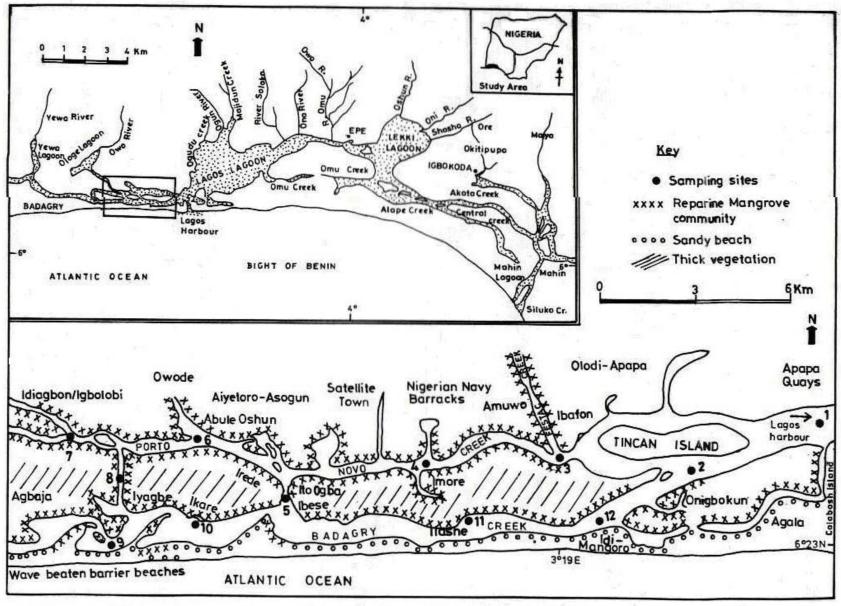


Fig. 1 lyagbe Lagoon, Porto-Novo and Badagry Creeks Showing Sampling Sites.



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#### **RESULT**

# Summary of physical and chemical results.

Presented below, is the summary of physical and chemical characteristics results for the lyagbe lagoon and for the duration of the study (Onyema and Nwankwo, 2009). Whereas air temperature, surface water temperature, total dissolved solids, transparency, sulphate, silica, dissolved oxygen, conductivity, salinity, chloride, pH, acidity, alkalinity, total hardness, calcium and magnesium recorded increasing values in the dry season, chemical oxygen demand, biological oxygen demand, total suspended solids, nitrate, phosphates, copper, zinc and iron recorded higher values in the wet seasons at all stations.

Table 2: A summary of the minimum, maximum and mean estimated values for environmental factors from the lyagbe lagoon (October, 2004 – September, 2006).

	Parameter / Unit	Minimum value	Maximum value	Mean value ± S.D.
1	Air temperature (°C)	26	34	30.07 ± 1.98
2	Water temperature (°C)	26	33	29.42 ± 1.81
3	Transparency (cm)	22	231	102.42 ± 51.47
4	Total Dissolved Solids (mgL <sup>-1</sup> )	90	25000	8467.65 ± 6641.66
5	Total Suspended Solids (mgL <sup>-1</sup> )	18	2310	172.48 ± 259.01
6	Rainfall (mm)	6	315.7	141.83 ± 116.87
7	Chloride (mgl <sup>-1</sup> )	20.5	15015	6316.55 ± 24167.13
8	Total hardness (mgL <sup>-1</sup> )	18	6875	2035.82 ± 1485.42
9	рН	6.7	8.42	7.40 ± 0.28
10	Conductivity (µS/cm)	110	40850	13208.59 ± 10418.71
11	Salinity (‰)	0.06	35.1	14.43 ± 18.10
12	Alkalinity (mgL <sup>-1</sup> )	15.3	330	74.32 ± 74.25
13	Acidity (mgL <sup>-1</sup> )	3.8	44	11.80 ± 7.48
14	Dissolved oxygen (mgL <sup>-1</sup> )	4	5.6	4.67 ± 0.23
15	Biological oxygen demand (mgL <sup>-1</sup> )	2	22	7.15 ± 3.52
16	Chemical oxygen demand (mgL <sup>-1</sup> )	8	89	30.21 ± 21.08
17	Nitrate – nitrogen (mgL <sup>-1</sup> )	3.3	59.8	10.54 ± 8.37
18	Phosphate – phosphorus (mgL <sup>-1</sup> )	0.01	1.68	0.26 ± 0.29
19	Sulphate (mgL <sup>-1</sup> )	20.8	1140	279.71 ± 232.16
20	Silica (mgL <sup>-1</sup> )	0.9	6.0	2.63 ± 0.91
21	Calcium (mgL <sup>-1</sup> )	10	720.1	188.49 ± 130.05
22	Magnesium (mgL <sup>-1</sup> )	1.4	900	333.36 ± 264.92
23	Copper (mgL <sup>-1</sup> )	0.001	0.079	0.003 ± 0.001
24	Iron (mgL <sup>-1</sup> )	0.06	1.08	0.29 ± 0.25
25	Zinc (mgL <sup>-1</sup> )	0.001	0.015	0.002 ± 0.002
26	Chlorophyll $a$ (µg/L)	4.2	55	19.63 ± 7.90

## **Phytoplankton**

The wet season algal community was dominated by freshwater species especially in more inland areas from the sea. The green algae, chrysophytes, euglenoids and pennate diatoms were prevalent in the wet season plankton, whereas dinoflagellates and most centric diatoms dominated the dry season. However the genus *Aulacoseira* with a number of varieties (centric diatoms) were

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reported only in the wet season / low salinity periods in the lagoon. Notable freshwater condition indicator included Aulacoseira granulata, Aulacoseira granulata var. angustissima, Aulacoseira granulata var. angustissima f. spiralis, Microcystis aureginosa and Microcystis flos — aquae. Similarly, mid to high brackish water / marine forms included Coscinodiscus centralis, Coscinodiscus eccentricus, Coscinodiscus radiatus, Coscinodiscus marginatus, Thalassionema frauenfeldii, Actinoptychus splendens, Melosira nummuloides, Melosira moniliformis, Bacillaria paxillifer, Parabelius delognei Pleurosigma and Gyrosigma spp. The dinoflagellates also fall into this latter category (Ceratium macroceros , Ceratium tripos and Peridinium africana).

Most pennate diatoms were prevalent in shallow eco-zones, whereas chrysophytes (*Chrysotepphanosphaera globulifera* and *Synura uvella*) reflected fresh and acidic water conditions and were only found in the furthest extreme of the lyagbe lagoon system and only in the Porto-Novo creek wing. The blue-green algae on the other hand (notably *Oscillatoria* spp.) were common in areas of fresh to low brackish water and moderate to high levels of nutrients, likely from organic pollution. *Merismopedia gluca* on the other hand was only recorded at times when mid to high brackish water situations were prevalent. Additionally, *Trichodesmium thiebautii* Gomont was recorded in sea conditions with low nutrient levels.

The euglenoids (*Euglena acus*, *Phacus curvicauda*, *Phacus acuminatus* and *Trachelomonas hispida*) were all found in the wet season associated with freshwater conditions and in areas with high nutrient levels throughout the study. Table 3 shows some elicited micro-algal bio-indicators of a range of water condition in the lyagbe lagoon for the study.

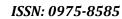
Table 3: Some recorded micro-algal bio-indicators of a range of water quality condition in the lyagbe lagoon.

	TAXA	PREFERENTIAL ECOLOGICAL SITUATION
	CLASS - CYANOPHYCEAE	
	ORDER I - CHROCCOCALES	
1	Chroococcus turgidus (Kutz.) Lemm.	Fresh – brackish water situation / moderate organic pollution
2	Microcystis aureginosa Kutzing	Fresh – low brackish / moderate – high organic pollution condition
3	Microcystis flos-aquae Kirchner	Fresh – low brackish / moderate – high organic pollution condition
4	Merismopedia gluca (Ehr.) Nageli	Mid – high brackish / moderate – high organic pollution condition
	ORDER 11– HORMOGONALES	
5	Anabaena constricta Geitler	Fresh water situation / moderate – high organic pollution condition
6	Anabaena spiroides Klebahn	Fresh – low brackish / moderate – high organic pollution condition
7	Anabaena torulosa Lagerheim	Fresh – low brackish / moderate – high organic pollution condition
8	Lynbgya limnetica Lemm	Fresh – low brackish / moderate organic pollution condition
9	Lynbgya martensiana Meneghini	Fresh – low brackish / moderate – high organic pollution condition
10	Oscillatoria borneti Zukal	Fresh – low brackish / high organic pollution condition
11	Oscillatoria chalybea Gomont	Fresh – low brackish / moderate – high organic pollution condition
12	Oscillatoria curviceps C.A. Agardh	Fresh water situation / moderate – high level organic pollution
13	Oscillatoria formosa Bory	Fresh water situation / moderate organic pollution
14	Oscillatoria limnosa Agardh	Fresh – low brackish / moderate – high organic pollution condition
15	Oscillatoria tenius Agardh	Fresh water situation / moderate organic pollution
16	Oscillatoria sancta Sancta	Fresh water situation / moderate organic pollution
17	Spirulina platensis Geitler	Fresh – low brackish / moderate – high organic pollution condition



18	Trichodesmium thiebautii Gomont	Low nutrient levels / warm sea situation / alkaline pH and high cation levels
$\vdash \vdash \vdash$	CLASS FUCLENOSINOSAS	
$\vdash$	CLASS – EUGLENOPHYCEAE ORDER – EUGLENALES	
19	Euglena acus Ehrenberg	Frach water situation / years high nutrient levels / organic nellution
20	Phacus curvicauda Swirenko	Fresh water situation / very high nutrient levels / organic pollution  Fresh water situation / very high nutrient levels / organic pollution
21		
-	Phacus acuminatus Stokes	Fresh water situation / very high nutrient levels / organic pollution
22	Trachelomonas hispida (Perry) Stein	Fresh water situation / very high nutrient levels / organic pollution
	CLASS - BACILLARIOPHYCEAE	
	ORDER 1 – CENTRALES	
23	Actinoptychus splendens Ehrenberg	Low brackish – sea situations / alkaline pH, high cation levels
24	Aulacoseira granulata Ehrenberg (Ralfs)	Moderate nutrient levels / fresh water – low brackish water situation
25	Aulacoseira granulata var. angstissima Muller	Moderate nutrient levels / fresh water – low brackish water situation
26	Aulacoseira granulata var. angstissima f. spiralis Muller	Fresh water situation / moderate level nutrient levels / moderate organic pollution
27	Aulacoseira granulata var. angstissima f. curvata Simon	Fresh water situation / moderate level nutrient levels / moderate organic pollution
28	Aulacoseira islandica (O.F. Muller) Simonson	Fresh water situation / moderate level nutrient levels
29	Melosira moniliformis Agardh	High brackish – sea situations / alkaline pH, high cation and nutrient levels
30	Melosira nummuloides Agardh	High brackish – sea situations / alkaline pH, high cation and nutrient levels
31	Ditylum brightwelli (T. West) Grunow	High brackish – sea situations / alkaline pH, high cation levels
32	Hemidiscus cuneiformis Wallich	High brackish – sea situations / alkaline pH, high cation levels
33	Cyclotella menighiniana Kutzing	High nutrient level / fresh – low brackish situations / shallow ecozone/ organic
	_	pollution
34	Leptocylindricus danicus Cleve	Low brackish – sea situations
35	Chaetoceros convolutus Castracane	Low brackish – sea situations / alkaline pH, high cation levels
	ORDER II – PENNALES	
36	Bacillaria paxillifer (O. F. Muller)	Mid – high brackish water situation / alkaline pH, high cation and nutrient levels
37	Eunotia monodon Ehrenberg	Fresh / acidic water situation
38	Fragillaria construens Ehrenberg	Fresh – low brackish / moderate – high level nutrient levels/ organic pollution
39	Gomphonema parvulum Grunner	Fresh – low brackish situations / High nutrient level / organic pollution
40	Gyrosigma balticum (Ehr.) Rabenhorst	Low – high brackish water situation / alkaline pH, high cation and nutrient levels.
41	<i>Gyrosigma spenceri</i> Wm. Smith	Low – high brackish water situation / alkaline pH, high cation and nutrient levels / shallow aqua-zone
42	Gyrosigma scalproides (Rabh) Cleve	Low – high brackish water situation / alkaline pH, high cation and nutrient levels
43	Hantzschia amphioxys (Ehr.) Rbenhorst	High brackish - sea water situation / alkaline pH, high cation and nutrient levels
44	Navicula cryptocephala (Kutz.) Hustedt	Moderate – high level nutrient levels / shallow aqua-zone / organic pollution
45	Navicula mutica Kutzing	Moderate – high level nutrient levels / shallow aqua-zone/ organic pollution
46	Navicula rhynchocephala Kutzing	High nutrient level / organic pollution / fresh – low brackish situations /
47	Nitzschia closterium Wm. Smith	Low – high brackish water situation/ alkaline pH and high cation levels
48		
	Nitzschia obtusa Wm. Smith	Low – high brackish / low nutrient water situation.
	Nitzschia obtusa Wm. Smith Nitzschia palea (Kutzing) Wm smith	Low – high brackish / low nutrient water situation.  High nutrient level / organic pollution / fresh situations
49 50	Nitzschia obtusa Wm. Smith Nitzschia palea (Kutzing) Wm smith Nitzschia sigmoidea (Witesch) Wm.	Low – high brackish / low nutrient water situation.  High nutrient level / organic pollution / fresh situations  Low – high brackish / high cation and nutrient situation.
49	Nitzschia palea (Kutzing) Wm smith	High nutrient level / organic pollution / fresh situations
49	Nitzschia palea (Kutzing) Wm smith Nitzschia sigmoidea (Witesch) Wm.	High nutrient level / organic pollution / fresh situations
49 50	Nitzschia palea (Kutzing) Wm smith Nitzschia sigmoidea (Witesch) Wm. Smith	High nutrient level / organic pollution / fresh situations  Low – high brackish / high cation and nutrient situation.
49 50 51	Nitzschia palea (Kutzing) Wm smith Nitzschia sigmoidea (Witesch) Wm. Smith Odontella regia (Schultze) Ostenfeld	High nutrient level / organic pollution / fresh situations  Low – high brackish / high cation and nutrient situation.  Mid – high brackish water situation / alkaline pH, high cation and nutrient levels

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	Smith	
55	Pinnularia major (Kutzing) Rabenh	Fresh water situation / moderate – high level nutrient levels/ organic pollution
56	<i>Pinnularia gibba</i> Ehrenberg	Fresh water situation / moderate – high level nutrient levels/ organic pollution / shallow area
57	Synedra ulna (Nitzsch) Ehrenberg	Fresh water situation / low - moderate level nutrient levels/ moderate organic pollution
58	Synedra ulna var. biceps Ehrenberg	Moderate – high level nutrient levels / Fresh – low brackish
59	Surirella ovata Kutzing	Shallow nutrient rich / Fresh – low brackish conditions/ moderate organic pollution
60	Surirella splendida Wm. Smith	Shallow nutrient rich / Fresh – low brackish conditions/ moderate organic pollution
61	Surirella striatula Turpin	Shallow nutrient rich / Fresh – low brackish conditions/ moderate organic pollution
	CLASS – DINOPHYCEAE	
	ORDER – PERIDINALES	
62	Ceratium macroceros (Ehr.) Cleve	Warm sea situations / alkaline pH
63	Ceratium tripos (O.F.M.) Nitzsch	Warm sea situations / alkaline pH
64	Peridinium africana Kofoid	Fresh – low brackish conditions / shallow aqua-zone
	CLASS – CHLOROPHYCEAE	
	ORDER I – ULOTHRICALES	
65	Microspora flocca (Vaucher) Thuret	High nutrient level / fresh situations / moderate organic pollution
66	Spirogyra africana Fritsch Cruda	High nutrient level / fresh situations / organic pollution
	ORDER II - CLADOPHORALES	
67	Cladophora glomerata (L) Kutzing	High nutrient level / fresh situations / organic pollution
	ORDER III - CHLOROCOCCALES	
68	Akistrodesmus sp.	Fresh / acidic water situation / shallow aqua-zone
69	Scenedesmus obliquus (Turp.) Kutzing	High nutrient level / fresh situations / organic pollution
70	Scenedesmus quadriquada (Turp.) de Brebisson	High nutrient level / fresh situations / organic pollution
	ORDER IV - ZYGNEMATALES	
71	Closterium ehrenbergii Meneghini	Fresh / acidic to neutral water pH / organic pollution
72	Gonatozygon monotaenium De Bary	Fresh / acidic to neutral water pH / moderate organic pollution
73	Gonatozygon sp.	Fresh / acidic to neutral water pH / moderate organic pollution
74	Staurastrum paradoxum var. cingulum Wm. and G.S. West	Fresh / acidic to neutral water pH / organic pollution
	CLASS – CHRYSOPHYCEAE	
75	Chrysotepphanosphaera globulifera Scherffel	Fresh / acidic water situation / shallow aqua-zone
76	Synura uvella Ehrenberg	Fresh / acidic water situation / shallow aqua-zone



#### **DISCUSSSION**

According to Onyema and Nwankwo (2009), the water quality indices in the lyagbe lagoon reflected seasonal changes closely related to rainfall distributive pattern and the effect of tidal seawater incursion. In the lagoon system, there were clear differences in phytoplankton species in the fresh, brackish and sea-like zones especially in the dry season. This corresponded with differences in the water quality.

The occurrence of pennate diatoms in the plankton during the survey may be reflections of possible stirring of the lagoon phytobenthic community into the plankton. According to Onyema et. al. (2003) frequently occurring pennate forms in the plankton samples from the Lagos lagoon was a likely reflection of the mixing of the shallow lagoon and phytobenthic community by tides and flood waters at different seasons. Nwankwo and Akinsoji (1989), Onyema and Nwankwo (2006) and Onyema et. al. (2007) are also of similar views for studies in the Lagos lagoon. According to Nwankwo (1984), the occurrence of pennate forms during the rainy season suggests their dislodgement from the substratum probably during high water discharge, while tidal inflow accounted for the appearance of some marine forms in the plankton at the same period. In this regard, the presence of known marine forms like Amphora alata, Asterionella japonica, Ditylum brightwellii, Melosira moniliformis, M. nummuloides, Triceratium favus and the various species of Coscinodiscus, Odontella, Chaetoceros, Rhizosolenia, Leptocylindricus, Thalassosira and Thalassionema further confirms the incursion of seawater to the lagoon as they were recorded in the mid-high brackish water situation only (Nwankwo and Onyema, 2003; Onyema et al., 2008).

According to Kadiri (1999) *Actinoptychus splendens, Aulacoseira granulata* and *Aulacoseira granulata* var. *angustissima* f. *curvata* had a wide distribution. The author is of the view that the prominence of *A. granulata* in all Nigerian coastal states, adds to the impressive array of evidence attesting to the cosmopolitan nature of this species. Its prevalence has been reported in other southern waters in Nigeria – (Lagos lagoon - Fox 1957, Nwankwo, 1988, 1996, Kadiri, 1999; Onyema *et al.* 2003; Onyema, 2008; Eleiyele reservoir - Imevbore, 1968; River Niger - Eaton, 1966; River Oshun - Egborge, 1973, 1974; Warri River - Opute, 1990).

Oscillatoria spp. according to Vanlandingham (1982) is by far the most significant of all bluegreen algae genera in determining water quality and it is highly important as a diagnostic indicator among other major algal groups. Palmer (1969) is of the view that only Euglena is more important than Oscillatoria as a genus of algae indicative of organic pollution. Desikachay (1959), Epstein (1995), Hallegraeff et. al. (1995) and Nwankwo et. al. (2003) have reporded that Trichodesmium thiebautii is commonly found in the supra thermocline, nutrient poor and warm waters of the tropics. Further to this, Dugale et. al. (1964) reporting for the Arabian sea and Nwankwo et. al. (2003) reporting for the Nigerian coast has reported massive blooms of Trichodesmium thiebautii clogging fishing nets and reducing fish catch. The presence of Trichodesmium thiebautii, the only true marine cyanobacteria also confirm the water chemistry at the time (Nwankwo, 1993; Nwankwo and Onyema, 2003; Onyema et al., 2008). Its source to the lagoon in definetly the sea (Atlantic ocean) via the Lagos habour.



According to Siver (2003), chrysophytes especially the synurophyceae are euplanktonic in nature and occur almost exclusively in freshwater habitat. The most diverse floras of scaled chrysophytes are typically found in slightly acidic condition. Chrysophytes encountered in this study were found only in such conditions which were also fresh at the time of occurence. Wujek *et al.* (2004) have recently listed new records of chrysophytes for Nigeria from the Lekki lagoon and this included *Synura uvella* encounted in this study but not *Chrysotepphanosphaera globulifera*.

With regard to diatoms, whereas *Coscinodiscus*, *Odontella* and *Parabelius* spp. and the dinoflagallates were more biodiganostic of marine condition and the dry season in the Iyagbe Iagoon, *Aulacoseira granulata*, *Aulacoseira granulata* var. *angustissima*, *Aulacoseira granulata* var. *angustissima* form *curvata*, *Aulacoseira granulata* var. *angustissima* form *spiralis*, *Microcystis aureginosa* and euglenoids better represented fresher water conditions in the Iyagbe Iagoon. According to Onyema (2007b), the community of *Biddulphia sinensis*, *Beggiotoa alba*, *Microcyctis flos* – *aquae*, *Spirulina platensis* and *Phormidium uncinatum* recorded in high numbers at one time or the other in the polluted Ijora creek were clear indicators of the alkaline, nutrient rich and brackish nature of the creek.

Aulacoseira granulata var. angustissima and Microcystis aeruginosa bloom in the wet season may be indication of a number of situations. Firstly, that the levels of nutrient in the waters are high. Secondly other associated hydro-climatic characteristics at such times may exist in favourable levels or state. Thirdly, these waters are very likely fresh as these organisms have been reported by other authors in coastal water of Nigeria as prevalent in freshwater situations hence indicative of fresh water conditions (Nwankwo, 1988, Kadiri, 1999, Nwankwo et al. 2003, Onyema et al. 2003). It is also possible that these organisms had bloomed in the adjoining Elete creek, Badagry, Yewa and Ologe lagoons and as a result of the rains and associated flood, overflows into the western extreme of the lyagbe lagoon. Nwankwo (1998) and Onyema et al. (2003) are of the view that high densities of Aulacoseira granulata and Aulacoseira granulata var. augustissima recorded in the Lagos lagoon in the wet season were possibly recruited from the eastern extremes of the Lagos lagoon system known to be fresh all through the year.

It is important to note that this study is not near exhaustive nor is the bio-diagonistic list presented in this material sacrosanct. Hence the need to continue to elucidate relevant biological data from ecological investigations in a bid to better understand phytoplankton species vis-à-vis the environmental conditions they portend. Apt identification in this regard is therefore very key.

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#### **REFERENCES**

- [1] Bettrons, D.A.S. and Castrejon, E.S. (1999). Biotropica. 31(1): 48 70.
- [2] Desikachary, T.V. (1959). Cyanophyta. Indian Council of Agric. Research, New Delhi, 686pp.
- [3] Dugale, R.C., Goering, J.J. and Rulther, J.H. (1964). High nitrogen fixation rates in the Sargasso

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- sea and the Arabian sea Limnol oceanoer. 9: 507-510.
- [4] Eaton, S. (1966). Ecological Studies of Phytoplankton in the newly forming Volta Lake of Ghana. Journal of West African Science Association. 11: 14 20.
- [5] Egborge, A.B.M. (1973). A preliminary checklist of the phytoplankton of Oshun River, Nigeria.
- a. Freshwater Biology. 4: 569 572.
- [6] Egborge, A.B.M. (1974). The seasonal variation and distribution of phytoplankton of Oshun River,
- a. Nigeria. Freshwater Biology. 4: 177 191.
- [7] Epstein, P.R. (1995). The role of algal blooms in the spread and persistence of human cholera. In:
- a. Harmful marine algal blooms. (Lassus P, Arzul, G., Erard-Le Denn, Gentien P. and Marcoillou-le Baut, C. (Eds.) p.846 850. Proceedings of the sixth International Conference on Toxic Marine Phytoplankton, October 1993, Nanter, France.
- [8] Fox, M. (1957). A first list of marine algae from Nigeria. Journal of Limnological Society of Botany London. LV(365): 615-631.
- [9] Hallegraeff, G.M., Anderson D.M. and A.D. Cembella (Eds.) (1995). Manual on Harmful marine microalgae. I.O.C. Manual and Guides 33 UNESCO France .551pp.
- [10] Hendey, N.I. (1958). Marine diatoms from West African Ports. Journal of Royal Microscopic Society. 77: 28-88.
- [11] Hendey, N.I. (1964). An introductory account of the smaller algae of British coastal waters. Part 5. Bacillariophyceae (diatoms) London. N.M.S.O. 317pp.
- [12] Imevbore, A.M.A. (1968). Planktonic algae of Eleiyele Reservoir, Nigeria. Journal of Science, Nigeria. 2: 85-90.
- [13] Kadiri, M.O. (1999). Phytoplankton distribution in some coastal waters of Nigeria. Nigeria Journal of Botany.12 (1): 51 62.
- [14] Nwankwo, D.I. (1984). Seasonal changes of phytoplankton of Lagos lagoon and the adjacent sea in relation to environmental factors. Ph.D. Thesis, University of Lagos. 447pp.
- [15] Nwankwo, D.I. (1986). Phytoplankton of a sewage disposal site in Lagos lagoon, Nigeria 1. The algae. Nigerian Journal of Biological Sciences. 1: 89-91.
- [16] Nwankwo, D.I. (1988). A preliminary checklist of planktonic algae in Lagos lagoon Nigeria. Nigeria. Journal of Botanica I Applied Sciences.. 2: 73-85.
- [17] Nwankwo, D.I. (1990). Contribution to the Diatom flora of Nigeria. Diatoms of Lagos lagoon and the adjacent sea. Nigerian Journal of Botany. 3: 53-70.
- [18] Nwankwo, D.I. (1993). Cyanobacteria bloom species in coastal waters of South-Western Nigeria. Archiv Hydrobiologie Supplement. 90: 553-542.
- [19] Nwankwo, D.I. (1996). Phytoplankton diversity and succession in Lagos Iagoon, Nigeria. Archiv
- a. Fur Hydrobiologie. 135(4): 529-542.
- [20] Nwankwo, D.I. (1998). The influence of sawmill wood wastes on Diatom population at Okobaba Lagos, Nigeria. Nigeria Journal of Botany. 11: 16-24.
- [21] Nwankwo, D.I. (2004a). A Practical Guide to the study of algae. JAS Publishers, Lagos. Nigeria. 84pp.
- [22] Nwankwo, D.I. (2004b). The Microalgae: Our indispensable allies in aquatic monitoring and biodiversity sustainability. University of Lagos Press. Inaugural lecture seris. 44pp.



- [23] Nwankwo, D.I. and Akinsoji, A. (1989). The Benthic Algal Community of a Sawdust Deposition Site in Lagos Lagoon. International Journal of Ecology and Environmental Sciences. 15: 197-204.
- [24] Nwankwo, D.I. and Akinsoji, A. (1992). Epiphyte community of water hyacinth, Eichhornia crassipes (MART) Solms in coastal waters of South Western Nigeria. Archiv fur Hydrobiologie. 124(4): 501-511.
- [25] Nwankwo, D.I. and Onyema., I.C. (2003). A checklist of planktonic algae off Lagos coast. Journal of Scientific Research Development.9: 75 82.
- [26] Nwankwo, D.I., Onyema, I.C. and Adesalu, T.A. (2003). A survey of harmful algae in coastal waters of south-western Nigeria. Journal of Nigerian Environmental Society. 1(2): 241 246.
- [27] Odiete, W.O. (1999). Environmental Physiology of Animals and Pollution. Diversified Resources Ltd., Lagos. 261pp.
- [28] Onyema, I.C. (2007a). Mudflat microalgae of a tropical bay in Lagos, Nigeria. Asian Journal of Microbiology, Biotechnology and Environmental Sciences. 9 (4): 877 883.
- [29] Onyema, I.C. (2007b). The phytoplankton composition, abundance and temporal variation of a polluted estuarine creek in Lagos, Nigeria. Turkish Journal of Fisheries and Aquatic Sciences. 7: 89 96.
- [30] Onyema, I.C. (2008). A checklist of phytoplankton species of the lyagbe lagoon, Lagos. Journal of Fisheries and Aquatic Sciences. 3(3): 167 175.
- [31] Onyema, I.C. (2009a). Notes on the existence of an additional lagoon in South-western Nigeria: Apese Lagoon. Journal of American Science. 5(4):151-156.
- [32] Onyema, I.C. (2009b). The Water Chemistry, Phytoplankton Biomass (Chlorophyll a), Episammic and Periphytic Algae of the Apese Lagoon, Lagos. Report and Opinion. 1(5): 31 40.
- [33] Onyema, I.C. (2009c). Pollution and the ecology of coastal waters of Nigeria. Dolps and Bolps Investment Limited, Lagos, Nigeria. 216pp.
- [34] Onyema, I.C. (2013a). Primary Production And Nutrients In An Open Tropical Lagoon. Nature and Science.11(3):102-106.
- [35] Onyema, I.C. (2013b). The Physico-Chemical Characteristics And Phytoplankton of the Onijedi Lagoon Lagos. Nature and Science. 11(1):127-135.
- [36] Onyema, I.C. and Nwankwo, D.I. (2006). The epipelic assemblage of a polluted estuarine creek in Lagos, Nigeria. Pollution Research. 25 (3): 459 468.
- [37] Onyema, I.C. and Nwankwo, D.I. (2009). Chlorophyll a dynamics and environmental factors in a tropical estuarine lagoon. Academia Arena. 1(1): 18 30.
- [38] Onyema, I.C., Otudeko, O.G. and Nwankwo, D.I. (2003). The distribution and composition of plankton around a sewage disposal site at Iddo, Nigeria. Journal of Scientific Research Development.7: 11-26.
- [39] Onyema, I.C., Nwankwo, D.I., Owolabi, K.O. (2008). Environment and Conservation. 14(4): 1 9.
- [40] Onyema, 1.C., Okpara, C.U., Ogbebor, C.I. Otudeko, O. and Nwankwo, D.I. (2007). Ecology, Environment and Conservation. 13: 1 12.
- [41] Opute, F.I. (1990). Hydrobiologia. 208: 101 109.
- [42] Palmer, M.C. (1969). Journal of Phycology. 5: 78-82.



- [43] Patrick, R. and Reimer, C.W. (1966). Monogr. Acad. Nat. Sci. Philadelphia. 686pp.
- [44] Patrick, R. and Reimer, C.W. (1975). Monogr. Acad. Nat. Sci. Philadelphia. 213pp.
- [45] Rosowski, J.R. (2003). Wehr, J.D. and Sheath, R.G. (Eds). Academic Press, New York. pp 383 422.
- [46] Siver, P.A. (2003). Wehr, J.D. and Sheath, R.G. (Eds). Academic Press, New York. pp 523 558.
- [47] Smith, G.M. (1950). McGraw-Hill, London.719pp
- [48] Stanier, R.Y. (1963). Edmondson, W.T. (Ed.). John Wiley and Sons, Inc. New York. 1248pp.
- [49] Vanlandingham, S.L. (1982). U.S. Environmental Protection Agency, EPA 60.
- [50] Wimpenny, R.S. (1966). The plankton of the sea. Faber and Faber Limited, London. 426pp.
- [51] Whitford, L.A. and Schmacher, G.H. (1973). A manual of freshwater algae. Sparks press Raeigh. 324pp.
- [52] Wujek, D.E., Adesalu, T.A. and Nwankwo, D.I. (2004). Tropical Freshwater Biology. 12/13: 99 103.